

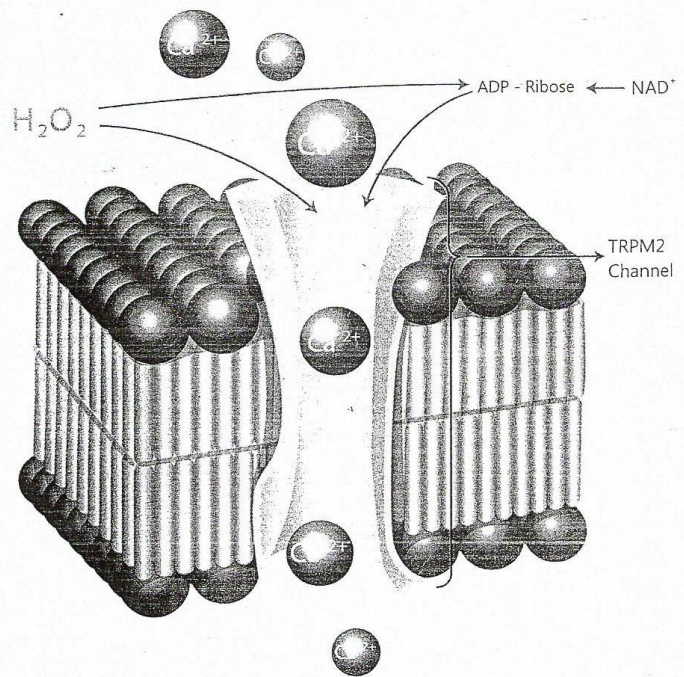
ISSN-1308-416X

# Cell Membranes and Free Radical Research

Volume 2 Number 1 1 June 2010

EDITOR-IN-CHIEF

Mustafa Naziroğlu, Isparta, TURKEY





---

Abstract Book  
of  
3<sup>rd</sup> International Congress on Cell  
Membranes and Oxidative Stress: Focus  
on Calcium Signaling and TRP Channels  
22-27 June 2010  
Isparta, Turkey

by  
Süleyman Demirel University Medical Faculty Department of Biophysics

---

# Cell Membranes and Free Radical Research

Volume 2 Number 1 1 June 2010

ISSN Numbers: 1308-4178 (On-line), 1308-416X

Indexing: Google Scholar, Index Copernicus, Chemical Abstracts, Scopus (Elsevier)

## EDITOR

### Editor In Chief

Mustafa Naziroglu. Department of Biophysics,  
Medical Faculty, Suleyman Demirel University, Isparta, Turkey.  
Phone: +90 246 211 33 10. Fax: +90 246 237 11 65  
E-mail: mnaziroglu@med.sdu.edu.tr

### Managing Editor

A. Cihangir Uğuz. Department of Biophysics, Medical Faculty,  
Suleyman Demirel University, Isparta, Turkey.  
E-mail: biophysics@med.sdu.edu.tr

## EDITORIAL BOARD

Cell Membranes, Ion Channels and Calcium Signaling  
Alexei Tepikin. The Physiological Laboratory, University of  
Liverpool, Liverpool, UK

Andreas Lückhoff. Institute of Physiology, Medical Faculty,  
RWTH-Aachen University, Germany.

Giorgio Aicardi. Department of Human and General Physiol-  
ogy, University of Bologna, Italy.

Jose Antonio Pariente. Department of Physiology, University of  
Extremadura, Badajoz, Spain.

James W. Putney, Jr. Laboratory of Signal Transduction,  
NIEHS, NC, USA.

Martyn Mahaut Smith, Department of Cell Physiology and  
Pharmacology, University of Leicester, Leicester, UK.

### Enzymatic Antioxidant Enzymes

Michael Davies. Deputy Director, The Heart Research Institute,  
Sydney, Australia.

Omer Akyol. Department of Biochemistry, Medical Faculty,  
Hacettepe University, Ankara, Turkey

Xingen G. Lei. Molecular Nutrition, Department of Animal Sci-  
ence, Cornell University, Ithaca, NY, USA

### Nonenzymatic Antioxidants, Nutrition and Melatonin

Ana B. Rodriguez Moratinos. Department of Physiology, Uni-  
versity of Extremadura, Badajoz, Spain.

Cem Ekmekcioglu. Department of Physiology, Faculty of Med-  
icine, University of Vienna, Austria.

Peter J. Battenworth. Nutritional Sciences Division, King's Col-  
lege, London, UK.

Şükrü Öter. Department of Physiology, GATA, Ankara, Turkey.

### Technical Editor

Ercan Sözbir. Department of Biophysics, Medical Faculty,  
Suleyman Demirel University, Isparta, Turkey.

E-mail: biophysics@med.sdu.edu.tr

İbrahim Kuş. Department of Graphic Design, Research  
Hospital, Suleyman Demirel University, Isparta, Turkey.

E-mail: ibrahim@sdu.edu.tr

## AIM AND SCOPE

Cell Membranes and Free Radical Research is a print and  
online journal that publishes original research articles, reviews  
and short reviews on the molecular basis of biophysical,  
physiological and pharmacological processes that regulate  
cellular function, and the control or alteration of these pro-  
cesses by the action of receptors, neurotransmitters, second  
messengers, cation, anions, drugs or disease.

Areas of particular interest are four topics. They are;

A- Ion Channels ( $\text{Na}^+$  -  $\text{K}^+$  Channels,  $\text{Cl}^-$  channels,  $\text{Ca}^{2+}$   
channels, ADP-Ribose and metabolism of  $\text{NAD}^+$ , Patch-  
Clamp applications),

B- Oxidative Stress (Antioxidant vitamins, antioxidant  
enzymes, metabolism of nitric oxide, oxidative stress, the  
biophysics of the radicals which spring up from oxygen ),

C- Interaction Between Oxidative Stress and Ion Channels  
(Effects of the oxidative stress on the activation of the voltage  
sensitive cation channels, effect of ADP-Ribose and  $\text{NAD}^+$  on  
activation of the cation channels which are sensitive to  
voltage, effect of the oxidative stress on activation of the TRP  
channels)

D- Gene and Oxidative Stress (Gene abnormalities. Inter-  
action between gene and free radicals. Gene anomalies and  
iron. Role of radiation and cancer on gene polymorphism)

## READERSHIP

Biophysics  
Biochemistry  
Biology  
Biomedical Engineering  
Pharmacology  
Physiology  
Genetics  
Cardiology  
Neurology  
Oncology  
Psychiatry  
Neuroscience

## KEYWORDS

Ion channels, cell biochemistry, biophysics, calcium signaling,  
cellular function, cellular physiology, metabolism, apoptosis,  
lipid peroxidation, nitric oxide synthase, ageing, antioxidants,  
neuropathy.



## *Organization Committee*

### *Congress Honor Committee*

Prof. Dr. Metin Lütfi BAYDAR,  
Rector of Süleyman Demirel University

Prof. Dr. Yıldırım SONGÜR, Dean of Faculty  
Medicine, Süleyman Demirel University

Prof. Dr. Süleyman KUTLUHAN,  
Chief Physician, Süleyman Demirel University  
Research Hospital

Prof. Dr. M. Salih ÇELİK,  
President of Turkish Biophysical Society

Aziz BAYRAK  
General Secretary of Süleyman Demirel University

### *Congress Organization Committee*

Prof. Dr. Mustafa NAZIROĞLU,  
Chairman  
Department of Biophysics, Faculty of Medicine,  
Süleyman Demirel University

Prof. Dr. James W. PUTNEY, Jr.  
Vice Chairman  
NIEHS Calcium Regulation Group Leader

Prof. Dr. Fatih GÜLTEKİN  
Department of Biochemistry, Faculty of Medicine  
Süleyman Demirel University

Assoc. Prof. Dr. Osman GÖKALP  
Department of Pharmacology, Faculty of Medicine  
Dicle University

### *Congress Secretariat*

A. Cihangir UĞUZ & İ. Suat ÖVEY &  
Bilal CİĞ  
Department of Biophysics, Faculty of Medicine  
Süleyman Demirel University

### *Accountants*

Ömer ÇELİK & Mustafa KÜÇÜKAYAZ &  
Cemil ÖZGÜL  
Department of Biophysics, Faculty of Medicine  
Süleyman Demirel University

### *Information Manager & Webmaster*

Ercan SÖZBİR  
Department of Biophysics, Faculty of Medicine  
Süleyman Demirel University

### *Technical Supports*

İns. Serdar Duran, İns. Hakan Mahmut  
Neğiş  
Department of Media & Public Relations  
Süleyman Demirel University



**TÜBİTAK**

*3<sup>rd</sup> International Congress on Cell Membranes and Oxidative Stress: Focus on Calcium Signaling and TRP Channels was supported by The Scientific and Technological Research Council of Turkey.*

*The Abstract book of the congress is published in this issue.*



Ram semen contains sufficient quantities of superoxide dismutase (SOD) and much lower concentrations of glutathione peroxidase (GSH-Px) and catalase (CAT) to prevent oxidative damage. The anti-oxidant capacity of the sperm cell is limited, due to a small cytoplasmic component, which contains these anti-oxidants to scavenge the oxidants. However, the concentration of these anti-oxidants may decrease considerably by the dilution of the semen. The aim of the present

#### Oral Presentation 9

### The influence of cysteine and taurine on microscopic-oxidative stress parameters and fertilizing ability of bull semen following cryopreservation

Serpil Sariözkan<sup>1</sup>, M.N. Bucak<sup>2</sup>, P.B. Tuncer<sup>2</sup>, P.A. Ulutaş<sup>3</sup>, A. Bilgen<sup>2</sup>

<sup>1</sup>Hakan Çetinsaya Experimental and Clinical Research Center, Faculty of Medicine, University of Erciyes, Kayseri, Turkey

<sup>2</sup>Ministry of Agriculture and Rural Affairs, Lalahan Livestock Central Research Institute, Ankara, Turkey

<sup>3</sup>Department of Biochemistry, Faculty of Veterinary Medicine, University of Adnan Menderes, Aydın, Turkey

Oxidative stress significantly damages sperm functions such as motility, functional integrity, endogenous antioxidant enzyme activities and fertility due to lipid peroxidation induced by reactive oxygen species (ROS). The aim of this study was to determine the effects of antioxidants such as taurine and cysteine in Bioxcell extender on standard semen parameters, fertilizing ability, lipid peroxidation (LPO) and antioxidant activities comprising reduced glutathione (GSH), glutathione peroxidase (GSH-Px), catalase (CAT) and superoxide dismutase (SOD) after the cryopreservation/thawing of bull semen. Nine ejaculates for each bull were included in the study. Three groups, namely taurine (2 mM), cysteine (2 mM), and control, were designed to analyze the antioxidants in Bioxcell. The addition of cysteine led to higher motility, compared to the other groups ( $p < 0.001$ ). Cysteine showed a greater protective effect on the percentages of acrosome and total abnormalities in comparison to the other groups ( $p < 0.001$ ). No significant differences were observed in hypo-osmotic swelling test (HOST), following supplementation with antioxidants during the freeze-thawing process. No significant difference was observed in non-return rates among groups. In biochemical assays, the additives did not show effectiveness on the elimination of malondialdehyde (MDA) formation and

#### 3rd International Congress on Cell Membranes and Oxidative Stress: Focus **2010** on Calcium Signaling and TRP Channels

maintenance of GSH and GSH-Px activities, when compared to controls. CAT activity was demonstrated to be significantly higher upon the addition of 2 mM taurine ( $p < 0.001$ ), while the level of MDA increased, indicating oxidative stress in this group. SOD activity was significantly elevated in the group with cysteine, compared to the other groups ( $p < 0.001$ ).

#### Oral Presentation 10

### The Effect of antioxidants on Sperm Parameters, Lipid peroxidation (LPO), Total glutathione (Total GSH) and Antioxidant Potential (AOP) Activities of Post-Thawed bovine Semen

methionine (2.5 and 7.5 mM), carnitine (2.5 and 7.5 mM), inositol (2.5 and 7.5 mM) and no additive (control), was cooled to 5°C and then frozen in 0.25 ml French straws. Frozen straws were thawed individually at 37°C for 20 sec in a water bath for the evaluation.

The extender supplemented with 7.5 mM doses of carnitine and inositol led to higher subjective motility percentages ( $61.9 \pm 1.3\%$  and  $51.3 \pm 1.6\%$ ) compared to the other groups. The addition of methionine and carnitine at doses of 2.5 and 7.5 mM and inositol at doses of 7.5 mM provided a greater protective effect in the percentages of total abnormality in comparison to the control and inositol 2.5 mM ( $p < 0.001$ ). In biochemical parameters, supplementation with antioxidants did not significantly affect LPO and total GSH levels in comparison to the control group ( $p > 0.05$ ). The