



Phytoplankton class determination by microscopic and HPLC-CHEMTAX analyses in the southern Baltic Sea

Elif Eker-Develi*, Jean-François Berthon, Dirk van der Linde

Joint Research Centre of the European Commission, Institute for Environment and Sustainability,
Global Environment Monitoring Unit, TP 272, 21027, Ispra (VA), Italy

ABSTRACT: The contribution of phytoplankton groups to total chlorophyll *a* (chl *a*) was derived using CHEMTAX from pigment measurements by HPLC and compared with the carbon (C) biomass estimations obtained from microscopy in the southern Baltic Sea in April 2005. Five different matrices of pigment:chl *a* input ratios, derived from the literature, were tested. Successive runs of CHEMTAX showed peridinin:chl *a* for dinoflagellates and fucoxanthin:chl *a* for diatoms to converge at 0.452 ± 0.02 (mean \pm SD) and 0.489 ± 0.03 , respectively, with initial ratios varying by a factor of 2 to 3 across matrices. The 2 techniques were in relatively good agreement for the dominant phytoplankton groups. Peridinin, diadinoxanthin, chlorophylls c_1 and c_2 (here grouped together as chl c_1+c_2), fucoxanthin and alloxanthin were the principal accessory pigments; dinoflagellates, diatoms and cryptophytes were the groups forming the majority of the C biomass. Diadinoxanthin and chl c_1+c_2 were mainly associated with the dominant dinoflagellates rather than with other phytoplankton classes. Excluding cyanophytes, the correlation between carbon biomass of other minor phytoplankton groups and their chl *a* was either poor or not significant due to uncertainties in either microscopic counts or CHEMTAX classification. There was a good correlation between carbon biomass of phytoplankton and chl *a*. The estimated C:chl *a* ratio of total phytoplankton varied between 8 and 40 (in average 20 ± 7), with a higher value for dinoflagellates (30 ± 17) than for diatoms (9 ± 7). Fucoxanthin-containing small flagellates might have led to the overestimation of the diatom contribution by CHEMTAX at a few stations.

KEY WORDS: CHEMTAX · Pigments · Phytoplankton carbon · C:chl *a* · Baltic Sea

Resale or republication not permitted without written consent of the publisher

INTRODUCTION

Quantifying the variations in phytoplankton biomass and taxonomic composition is relevant for the evaluation of ecosystem function and status, as well as for the study of global biogeochemical cycles. The retrieval of main phytoplankton groups from ocean colour satellite data is also an important input for primary production models operating at the global scale (Alvain et al. 2005, Uitz et al. 2006), as photosynthetic parameters may vary according to taxonomic composition. Traditionally, phytoplankton groups are quantified under light microscope by identifying and counting cells, measur-

ing their size, and converting the resulting volumes (V) to carbon (C) biomass on the basis of previously established equations (Strathmann 1967, Verity et al. 1992, Montagnes & Franklin 2001). However, depending on the equation chosen for this conversion, calculated C values may vary for a specific taxonomic group (Havskum et al. 2004, Llewellyn et al. 2005). Moreover, intra-class variations up to ~30-fold in the C:V ratio of phytoplankton were observed (Llewellyn et al. 2005 and references therein). A more recent method to estimate the distribution of phytoplankton groups is based on the measurement of pigment composition of cells making use of the HPLC technique (Mantoura &

*Email: elif.eker-develi@jrc.it

Llewellyn 1983, Rodriguez et al. 2002). This technique is faster and more appropriate for the identification of pico- and nano-phytoplankton than the microscope method as complementary to flow-cytometry (Veldhuis & Kraay 2004). Thus, HPLC has been increasingly used, in particular for the large-scale mapping of pigments and phytoplankton groups in the world oceans (Tester et al. 1995, Vidussi et al. 2001, Llewellyn et al. 2005, Wänstrand & Snoeijs 2006). Nevertheless, important uncertainties in retrieved phytoplankton composition may result from the variability of the cellular pigment contents due to light and temperature changes, nutrient availability, distinct growth phases, as well as from the preclusion of phycobiliprotein quantification through HPLC. Furthermore, the ambiguous character of some marker pigments (i.e. their being shared by several phytoplankton groups and even by ciliates) in addition to intra-class and inter-species variation of pigment:chlorophyll *a* (chl *a*) ratios, are other issues which have to be cautiously considered in pigment studies (Schlüter et al. 2000, Henriksen et al. 2002, Irigoien et al. 2004). Lastly, non-pigmented heterotrophic phytoplankton species cannot be monitored with the HPLC method.

In order to relate the measured pigment composition to the distribution of phytoplankton groups, statistical techniques like multiple linear regression (Tester et al. 1995) or matrix factorization, such as the well known CHEMTAX software (Mackey et al. 1996), can be used. However, simple multiple linear regression analysis has several shortcomings compared to CHEMTAX, such as the occurrence of negative contributions to total chl *a* (Henriksen et al. 2002 and references therein). CHEMTAX, which uses predefined pigment:chl *a* ratios, is a powerful and robust approach for the taxonomic classification of phytoplankton assemblages in spite of the limitations related to pigment investigations mentioned above (Mackey et al. 1996, Schlüter et al. 2000, Rodriguez et al. 2002).

The C:chl *a* ratio of phytoplankton is also highly variable and depends on light, temperature, nutrients and species composition. Values ranging between 10 (Laws & Bannister 1980) and 250 or more (Cullen 1982, Veldhuis & Kraay 2004) were reported for natural phytoplankton populations. Several studies reported a good agreement between C biomasses of phytoplankton groups (as derived from microscopic determination) and class-related chl *a* concentrations (as derived from CHEMTAX analysis) for large diatoms, but a poor conformity for dinoflagellates, prymnesiophytes and small flagellates (Llewellyn et al. 2005 and references therein). This relationship between phytoplankton pigments and C biomasses of the corresponding taxonomic groups should be further investigated as the results obtained from CHEMTAX analysis might

differ according to region, environmental characteristics, phytoplankton composition, or phytoplankton biomass.

The aim of the present work was to investigate the applicability of CHEMTAX to the southern Baltic Sea by using simultaneous microscopic determinations of phytoplankton biomass and HPLC pigment analyses. In particular, the use of the classical Mackey et al. (1996) pigment:chl *a* input ratios, initially adapted for the open ocean environment, was tested in a highly turbid coastal region, together with other sets of ratios. In this context, validation of a recent study relying on CHEMTAX simulations which allow the use of *a priori* non-appropriate input ratios (Latasa 2007), was performed. From CHEMTAX and cell-counting results, C:chl *a* ratios were derived for total phytoplankton as well as for main groups. These ratios were compared with environmental conditions and simulated ratios obtained from phytoplankton acclimation models (Geider et al. 1997, Blackford et al. 2004).

MATERIALS AND METHODS

Surface water sampling for pigment and phytoplankton analyses was performed at 31 stations in the southern Baltic Proper between 16 and 29 April 2005, onboard the RV 'Oceania' as a result of a collaboration between the Institute of Oceanography of the Polish Academy of Sciences (IOPAS) and the Joint Research Centre of the European Commission (JRC) (Fig. 1).

Physicochemical and bio-optical measurements. Vertical profiles of temperature and salinity were performed using a Seabird-19 CTD at a depth of ~30 m. The depth of the surface mixed layer (D_{ML}) was defined as the depth where a change of $>0.3^{\circ}C$ in temperature over a 5 m depth interval was observed (Llewellyn et al. 2005).

Underwater and surface radiometric measurements of downwelling irradiance at 7 wavelengths (from 412 to 685 nm) were performed with a MiniNess free-fall profiler and surface reference (Satlantic). Above-surface photosynthetically available radiation (PAR) values (in $\mu mol\ photons\ m^{-2}\ s^{-1}$) were derived from these measurements and are here referred to as 'irradiance' (I). Average I in the surface mixed layer (SML) was estimated using the equation of Llewellyn et al. (2005) as follows:

$$I_{SML} = I_S D_{50\%} / D_{ML} \quad (1)$$

where I_{SML} is average irradiance in SML, I_S the surface irradiance, and $D_{50\%}$ the depth at which irradiance was 50 % that of the surface. Optimal C:chl *a* ratios over the observed range of I_{SML} and the temperature response factor were derived from models of Blackford et al.

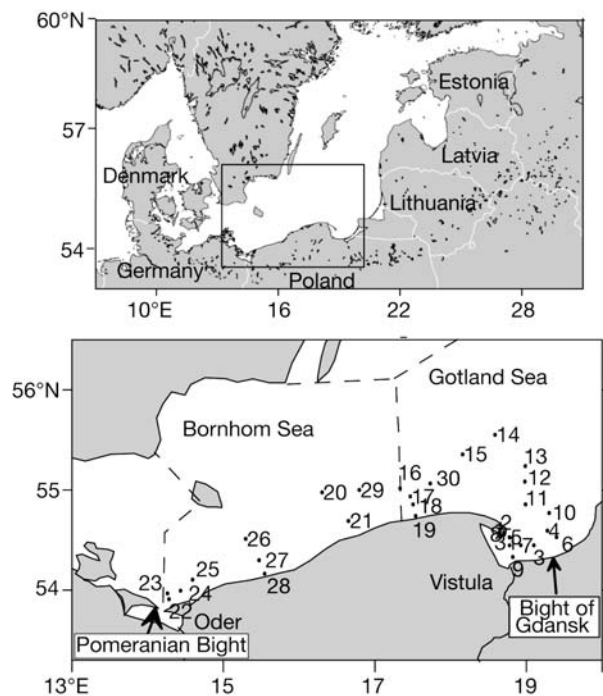


Fig. 1. Phytoplankton and pigment sampling stations in the southern Baltic Proper during April 2005

(2004), which were modified after Geider et al. (1997), assuming the proportion of photosynthate directed towards chlorophyll balances the existing C:chl *a* ratio. Initial slope of the photosynthesis-versus-irradiance (P-I) curve, light inhibition of photosynthesis, assimilation rate and maximum C:chl *a* ratio were given as $2.98 \text{ (W m}^{-2}\text{)}^{-1} \text{ d}^{-1}$, $0.02 \text{ (W m}^{-2}\text{)}^{-1} \text{ d}^{-1}$, 1 to 3 d^{-1} and 0.075, respectively in this model (Blackford et al. 2004, Llewellyn et al. 2005).

Phytoplankton analysis. Water samples for phytoplankton identification were put into 250 ml amber glass bottles to which a neutral lugol solution was added to obtain a final concentration of 1%. Note that lugol does not allow distinction between phototrophic and heterotrophic flagellates. For the preparation of samples for microscopic analysis, the sedimentation method was used (Eker-Develi et al. 2006a). Samples were concentrated to a *V* of ~40 ml and the micro- and nanophytoplankton were counted using a Sedgewick-Rafter counting chamber under a phase-contrast binocular microscope. Approximately 400 cells were counted in each sample. The identified species were classified under 8 phytoplankton taxonomic groups: dinoflagellates (Dino), diatoms (Diat), cryptophytes (Crypto), cyanophytes (Cyano), chlorophytes (Chloro), euglenophytes (Eugleno), raphidophytes (Raphido), chrysophytes (Chryso) and small flagellates (sFlag) (Table 1). A few rare prasinophyte species were included in the C biomass of the Chloro group. Chrysophytes did not include picoplanktonic

Table 1. Abbreviations used throughout this article for phytoplankton groups and photosynthetic pigments

	Abbreviation
Phytoplankton	
Dinoflagellates	Dino
Diatoms	Diat
Cryptophytes	Crypto
Cyanophytes	Cyano
Chlorophytes	Chloro
Euglenophytes	Eugleno
Raphidophytes	Raphido
Chrysophytes	Chryso
Small flagellates	sFlag
Pigment	
Chlorophyll <i>a</i>	Chl <i>a</i>
Peridinin	Peri
Diadinoxanthin	Ddx
Chlorophyll <i>c</i>	Chl <i>C</i> ₁₊₂
Fucoxanthin	Fuco
Alloxanthin	Allo
Chlorophyll <i>b</i>	Chl <i>b</i>
β-carotene	Caro
Diatoxanthin	Dtx
19'-butanoyloxyfucoxanthin	But
Zeaxanthin	Zea
19'-hexanoyloxyfucoxanthin	Hex
Chlorophyllide <i>a</i>	Chlide <i>a</i>

pelagophyceae. Biomass of the phototrophic ciliate *Mesodinium rubrum* (Lohmann) Jankowski (= *Myrionecta rubra*) was included in cryptophyte biomass as in previous studies by Henriksen et al. (2002) and Schlüter & Møhlenberg (2003).

The *V* of each cell was calculated by measuring its appropriate morphometric characteristics (i.e. diameter, length and width) (Kovalá & Larrance 1966, Olenina et al. 2006). One μm^3 *V* was assumed equivalent to 1 pg wet wt (Wasmund et al. 1998, Gasiunaite et al. 2005). For each identified cell, C biomass was calculated from the measured *V* using the equations of Strathmann (1967) for diatoms (Eq. 2) and other phytoplankton groups (Eq. 3) as follows:

$$\log C = -0.422 + 0.758 (\log V) \quad (2)$$

$$\log C = -0.46 + 0.866 (\log V) \quad (3)$$

C biomass of small phytoplankton for cell *V* (in μm^3) of 10^1 , 10^2 and 10^3 , was calculated by multiplying these *V* with density values (in $\text{pg C } \mu\text{m}^{-3}$) of 0.36, 0.24 and 0.16, respectively (Verity et al. 1992). The sFlag group may include chlorophytes, cryptophytes, chrysophytes and (less probably) cyanophytes (the majority of the cyanophytes were colony forming species in the present study) in the size range ~2 to 20 μm . C biomass of specific phytoplankton groups (here abbreviated as '[Group]-C') calculated according to Strathmann (1967) were compared with those obtained using equations established by other investigators. In the present

study, Diat-C calculated from the equations of Montagnes et al. (1994) and Montagnes & Franklin (2001) was higher than that calculated from Eq. (2) by an average factor of 2.1 (± 0.2) and 1.9 (± 0.4), respectively. The Dino-C found using the equations of Verity et al. (1992) and Menden-Deuer & Lessard (2000) were higher than values calculated by Eq. (3) by an average factor of 1.2 (± 0.05) while the Dino-C estimated from the equation of Montagnes et al. (1994) was lower by an average factor of 0.9 (± 0.1) than the C estimated by Eq. (3). When the equations given by Menden-Deuer & Lessard (2000) for chrysophytes (here applied to small flagellates too) and chlorophytes were used, the total C biomass of these groups decreased by an average factor of ~ 6 (5.5 ± 1.6) with respect to calculations using the equation of Strathmann (1967) which also includes values of Verity et al. (1992) for small phytoplankton noted above. The C biomass of *Mesodinium rubrum* was calculated with Eq. (3), as for cryptophytes. However, the C biomass of *Mesodinium* would double if the conversion factor of Putt & Stoecker (1989) for oligotrichous ciliates (0.19 pg pm^{-3}) were used.

Pigment analysis. The method used for the HPLC pigment analysis followed the core measurement protocols of the Joint Global Ocean Flux Study (JGOFS 1994), and is a modified version of the method presented by Wright et al. (1991). We collected 0.8 to 6 l of seawater and filtered it through 47 mm GF/F Whatman glass fibre filters. The filters were stored in liquid nitrogen for the duration of the cruise and then kept at -80°C until taken to the laboratory for analysis. Each filter was thawed and subsequently transferred into a plastic Falcon tube containing 3 ml of a solution of 100% acetone and an internal standard; it was then sonicated in a 0°C water bath for 1 min and left to extract for 24 h at -20°C . Extracts were clarified by filtering them onto $0.45 \mu\text{m}$ Teflon syringe filters, and subsequently injected into a Hewlett Packard/Agilent 1100 HPLC system. The HPLC system used includes a reversed-phase C_{18} column ($250 \times 4.6 \text{ mm}$, $5 \mu\text{m}$ particle size, Hichrom, with a C_{18} guard column), an autosampler with thermostat, a diode array detector (DAD), a pumping system and degasser, data acquisition and analysis software and a 3 solvent gradient. The 3 solvents were (1) methanol 80%–ammonium acetate (0.5 M) 20% (2) acetonitrile 90% and (3) ethyl acetate. The temperature-controlled autosampler (set at 4°C) mixed $75 \mu\text{l}$ of extract with $22.5 \mu\text{l}$ of distilled water buffer before injection through a $100 \mu\text{l}$ loop onto the C_{18} column. Chromatograms were generated from the absorbance of individual pigments at 436 nm for chlorophyll pigments and carotenoids, and at 405 nm for phaeopigments using DAD. The HPLC system was calibrated using pigment standards from DHI Water & Environment. This method does not permit the separa-

tion of divinyl chl *a* and chlorophyll *b* (chl *b*) from monovinyl chl *a* and chl *b*, respectively. It is not capable of fully separating chlorophyll c_1 and chlorophyll c_2 (presented here as chl c_1+c_2) either. $\beta\gamma$, $\beta\epsilon$ and $\beta\beta$ carotenes were not resolved well and their sum was given as β -carotene. The individual pigments measured were thus (Table 1): chl *a*, peridinin (Peri), diadinoxanthin (Ddx), chl c_1+c_2 , fucoxanthin (Fuco), alloxanthin (Allo), chl *b*, β -carotene (Caro), diatoxanthin (Dtx), 19'-butanoyloxyfucoxanthin (But), zeaxanthin (Zea), 19'-hexanoyloxyfucoxanthin (Hex) and the degradation product of chl *a*, chlorophyllide *a* (chl *a*).

A detailed description of this method and the associated uncertainties is presented in Hooker et al. (2000) and Claustre et al. (2004). Uncertainties on determination of pigment concentrations, defined as absolute percent differences between 4 laboratories (including JRC) were approximately 7% for the total chl *a* and 21.5% on average for the other pigments (ranging from 11.5% for fucoxanthin to 32.5% for peridinin).

CHEMTAX processing. The CHEMTAX program (Mackey et al. 1996) was used in order to retrieve the group composition of phytoplankton from some marker pigments. The 3 matrices that this method uses as inputs are (1) the measured concentration of marker pigments in the samples; (2) the theoretical ratios of marker pigments to chl *a* for each phytoplankton group to be quantified; and (3) a ratio limit matrix restricting the iterative adjustments of these ratios operated by CHEMTAX.

On the basis of the initial estimate of the pigment:chl *a* ratios for each selected group to be determined (Matrix 2), CHEMTAX optimizes the contribution of each phytoplankton group to the total chl *a* using a factor analysis and a steepest-descent algorithm and finds the best fit to the measured pigment data. Several initial accessory pigment:chl *a* ratio matrices were tested here (see below). Beside the partial chl *a* attributed to each phytoplankton group, one other output is a new matrix of pigment:chl *a* ratios resulting from the best fit. For each sample, 10 successive CHEMTAX runs were performed by using the output pigment:chl *a* ratios matrix of each run as input for the following run in order to have those ratios stabilize toward their most probable values (Latasa 2007). The parameters set for the calculations were as given by Latasa (2007).

The different input pigment:chl *a* ratio matrices tested here include or exclude chl c_1+c_2 , Dtx and Caro, which are shared by many phytoplankton groups (Jeffrey & Vesik 1997). Ddx was not taken out from any of the initial ratio matrices, as it was strongly correlated with the unshared pigment of dinoflagellates, Peri ($r^2 = 0.98$, $p < 0.05$). It should also be noted that Allo:chl *a* ratios used in the different matrices corresponded to

free-living cryptophytes, not to the kleptoplastidic ciliate *Mesodinium rubrum*. The following pigment:chl *a* ratio matrices were tested: (1) the mean values from Mackey et al. (1996) for 7 accessory pigments (Table 2, Appendix 1, available as MEPS Supplementary Material at www.int-res.com/articles/suppl/m359p069_app.pdf; NB chl *a* does not include chl *a*); (2) the values for chl c_1+c_2 , Dtx and Caro from Mackey et al. (1996), as well as the 7 accessory pigments in Table 2; (3) the mean values from Schlüter et al. (2000) for the 7 accessory pigments from Table 2 for species of mainly Scandinavian waters; (4) the same groups as in (3), but including the mean values for chl c_1+c_2 , Dtx and Caro taken from Schlüter et al. (2000); and (5) the maximum values for each phytoplankton group from Mackey et al. (1996), Schlüter et al. (2000) and Henriksen et al. (2002) for the same 7 accessory pigments of Table 2, in order to test the capacity of CHEMTAX to deal with extreme input ratios. However, for chrysophytes, But:chl *a* ratios of Mackey et al. (1996) were used in all input ratio matrices.

Model II (major axis) linear regression was used in order to assess the functional relationship between random variables (pigment and/or carbon concentrations). Note that the presented regression equations should not be used to predict values of *y* given *x* (Sokal & Rohlf 2000).

RESULTS

CHEMTAX-derived chl *a* estimations with different pigment input ratios

The variability of the final chl *a* allocation to the major groups (Diat and Dino) due to the use of different pigment ratio matrices was reduced by running the program 10 successive times as reported by Latasa (2007) (Table 3). Thus, the usage of distinct input pigment:chl *a* ratios did not result in significant differences (relative difference from the average was 1 to

Table 2. Input (from Mackey et al. 1996) and output (after 10 runs) ratios of marker pigments to chl *a* for the selected phytoplankton groups. Abbreviations defined in Table 1

	Marker pigments							
	Peri	But	Fuco	Allo	Ddx	Zea	chl <i>b</i>	chl <i>a</i>
Input ratios								
Dino	0.532	0	0	0	0.121	0	0	1
Diat	0	0	0.46	0	0.24	0	0	1
Crypto	0	0	0	0.23	0	0	0	1
Chryso	0	0.467	0.560	0	0.28	0	0	1
Eugleno	0	0	0	0	0.23	0	0.41	1
Chloro	0	0	0	0	0	0.059	0.285	1
Cyano	0	0	0	0	0	0.11	0	1
Output ratios								
Dino	0.424	0	0	0	0.181	0	0	1
Diat	0	0	0.465	0	0.059	0	0	1
Crypto	0	0	0	0.258	0	0	0	1
Chryso	0	0.465	0.560	0	0.279	0	0	1
Eugleno	0	0	0	0	7.945	0	9.838	1
Chloro	0	0	0	0	0	0.057	0.458	1
Cyano	0	0	0	0	0	0.770	0	1

Table 3. Chl *a* contributions of distinct phytoplankton groups to the total chl *a* calculated by CHEMTAX using different initial ratio matrices. Matrix 5 is the maximal pigment:chl *a* ratios for each phytoplankton group reported in Mackey et al. (1996), Schlüter et al. (2000) and Henriksen et al. (2002). Abbreviations defined in Table 1

Ratio matrix no.	Chl <i>a</i> contributions (%)							No. of pigments	Source
	Dino	Crypto	Chryso	Eugleno	Chloro	Cyano	Diat		
Output of 1st run									
1	44.34	33.57	2.06	0.27	1.52	1.01	17.23	7	Mackey et al. (1996)
2	40.44	36.55	1.36	3.53	0.99	2.17	14.96	10	Mackey et al. (1996)
3	31.11	35.36	1.32	1.05	1.36	15.81	14.00	7	Schlüter et al. (2000)
4	44.76	33.36	1.27	4.98	3.60	0.37	11.66	10	Schlüter et al. (2000)
5	55.11	2.19	0.41	2.23	19.91	0.25	19.91	7	
Avg	43.15	28.20	1.28	2.41	5.48	3.92	15.55		
SD	8.76	14.60	0.59	1.89	8.13	6.69	3.15		
Output after 10th run									
1	44.40	33.00	1.30	0.10	1.90	0.70	18.60	7	Mackey et al. (1996)
2	41.40	35.24	1.33	3.24	1.10	1.31	16.38	10	Mackey et al. (1996)
3	39.14	33.41	1.32	1.75	0.95	6.13	17.31	7	Schlüter et al. (2000)
4	41.88	32.31	1.30	5.59	2.04	0.40	16.46	10	Schlüter et al. (2000)
5	42.52	2.14	0.41	1.61	32.67	0.26	20.39	7	
Avg	41.87	27.22	1.13	2.46	7.73	1.76	17.83		
SD	1.90	14.06	0.41	2.08	13.95	2.47	1.69		

15%) in chl *a* allocations to these 2 groups (as seen in the results of the tenth run for Matrices 1 through 5 in Table 3). Even though Crypto is a major phytoplankton group, the Allo:chl *a* ratio did not converge when a very high ratio (~10 to 20 times higher than in other matrices) was used in Matrix 5. Successive runs of CHEMTAX did not bring about the convergence of pigment:chl *a* ratios; thus there was no convergence of chl *a* allocations for minor phytoplankton groups (Table 3). The percentage contributions of the minor groups Chryso, Chloro, Eugleno and Cyano to the total chl *a* differed (relative difference ~20 to 190%) among distinct ratio matrices.

Based on the comparison between phytoplankton C estimations and pigment concentrations, Matrix 1 was chosen as the most appropriate among the tested matrices, though it did not produce a good correlation between group-chl *a* and group-C for minor phytoplankton groups. Matrices 2 and 4 included the ambiguous pigments chl c_1+c_2 , Caro and Dtx, and both produced high contribution by Eugleno to the total chl *a* (Table 3). However, the C biomass of euglenophytes (which are easy to identify under microscope) was not so high in the samples compared to Chloro-C or Cyano-C. Considering the low Zea concentrations, the contribution of Cyano to the total chl *a* appeared to be overestimated upon running Matrix 3 (Table 3). When Matrix 5 was run, Chloro-chl *a* was found to be higher than Diat-chl *a* and Crypto-chl *a*, which does not seem possible, given that C biomass of diatoms and cryptophytes were ~20 and 70 times higher, respectively, than that of chlorophytes. Furthermore, Fuco and Allo concentrations were ~4 times higher than chl *b*.

The ratios Fuco:chl *a* for Diat and Peri:chl *a* for Dino converged (at 0.452 ± 0.02 and 0.489 ± 0.03 , respectively) at the end of the tenth run in all matrices used, even though initial input ratios were about 2 to 3 times higher for Matrix 5 than for the others (see Appendix 1, Fig. 2). The Allo:chl *a* ratio converged (at 0.251 ± 0.01) when Matrices 1 through 4 were used but (as mentioned above) remained constant in Matrix 5. Pigment:chl *a* ratios of minor groups did not converge. For comparisons of different phytoplankton groups' contributions to total C and chl *a*, results from only Matrix 1 are shown.

Hydrography

Surface temperature and salinity (average values within the first meter) showed a gradient, with values from ap-

proximately 9°C and 3.5 psu in the Bight of Gdańsk to between 4 and 5°C, and 7 and 8 psu in the open Baltic Sea. In the southern Baltic Sea, the maximum fresh water runoff occurs in April-May and coincides with the phytoplankton spring bloom initiated by the stabilization of the water column and increased surface light (Kowalczyk et al. 2005). The Vistula and Oder, flowing into the Gdańsk and the Pomeranian Bights, respectively, are the major rivers draining most of Poland.

Species and size composition of phytoplankton

Autotrophic dinoflagellate *Perdinella catenata* Levanter had the highest average C biomass among dinoflagellate species, accounting for 29% of total C biomass and 58% of Dino-C in the study region excluding 4 stations in the Pomeranian Bight where this species was not detected (Table 4). This species contributed to the total dinoflagellate C biomass at a higher rate in the Gotland Sea, accounting for 67% of total Dino-C. The second most important species in terms of C was the phototrophic ciliate *Mesodinium rubrum* Lohmann, which constituted 70% and 60% of total phytoplankton C biomass at the first 2 sampling stations. The diatom *Thalassiosira baltica* (Grunow) Ostefeld (Table 4), which was also more abundant in the Gotland Sea (60% of total Diat-C), was the third most important species in terms of C. Except *Asterionella formosa* Hassall, which was more abundant in the Bornholm Sea, other species noted in Table 4 also had higher C biomass in the Gotland Sea.

Phytoplankton cells 10 to 100 μm^3 in *V* constituted ~80% of total abundance, while ~70% and 20% of total C biomass were formed by cells 10³ to 10⁴ μm^3 and 10⁴ to 10⁵ μm^3 in *V*, respectively.

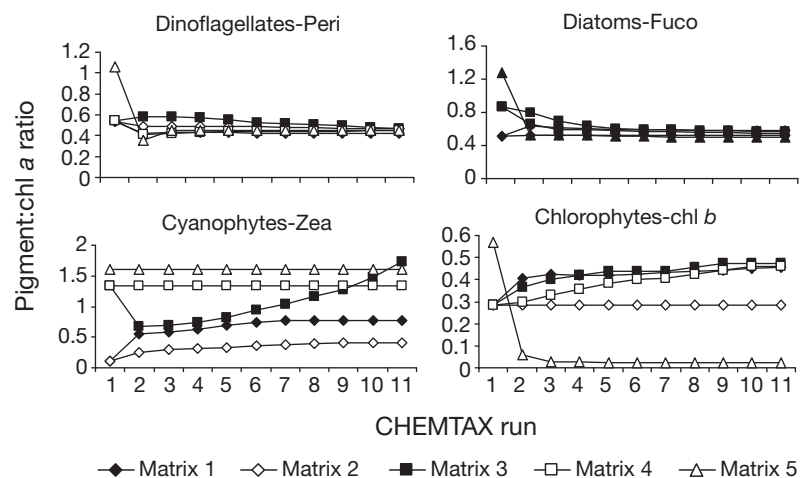


Fig. 2. Variations in pigment ratios after successive runs of CHEMTAX for 4 phytoplankton pigment groups. Abbreviations defined in Table 1

Table 4. Dominant phytoplankton species, their phototrophic characters (for dinoflagellates) and average carbon concentrations at the sampling stations

Group Species	Phototrophic character (source)	Average carbon ($\mu\text{g l}^{-1}$)
Dinoflagellate		
<i>Peridiniella catenata</i> Levander	Autotrophic, Braleswska & Witek (1995)	91 ± 160
<i>Achradina pulchra</i> Lohmann	No information	9 ± 11
<i>Gymnodinium</i> spp.		2 ± 10
<i>Heterocapsa rotundata</i> (Lohmann) comb. nov. (= <i>Katodinium rotundatum</i> (Lohmann) Loeblich)	Autotrophic, Braleswska & Witek (1995)	2 ± 4
<i>Gyrodinium spirale</i> (Bergh) Kofoid et Swezy	Heterotrophic, Kim & Jeong (2004)	2 ± 2
<i>Protoperidinium bipes</i> (Paulsen) Balech	Heterotrophic, Braleswska & Witek (1995)	1 ± 2
<i>Katodinium glaucum</i> (Lebour) Loeblich III	Heterotrophic or mixotrophic, Mouritsen & Richardson (2003)	1 ± 1
Ciliate		
<i>Mesodinium rubrum</i> Lohmann (= <i>Myrionecta rubra</i> Jankowski)	Photosynthetic, Gustafson et al. (2000)	70 ± 109
Diatom		
<i>Thalassiosira baltica</i> (Grunow) Ostensfeld	Autotrophic, Jeffrey & Vesk (1997)	9 ± 19
<i>Skeletonema costatum</i> (Grev.) Cleve		2 ± 4
<i>Asterionella formosa</i> Hassall		1 ± 3
<i>Coscinodiscus</i> sp.		1 ± 1
Cyanophyte		
<i>Aphanizomenon flos-aqua</i> (L.) Ralf	Autotrophic, Jeffrey & Vesk (1997)	3 ± 6
Cryptophyte		
<i>Teleaulax amphioxeia</i> (Conrad) Hill	Autotrophic, Janson (2004)	2 ± 2

Spatial distribution of Phyto-C and chl a

The C biomass of total phytoplankton (Phyto-C) and chl a (Fig. 3) were concordant with the salinity gradient. The highest Phyto-C and chl a were observed in the least saline region of the Bight of Gdańsk (more than $965 \mu\text{g C l}^{-1}$ and $50 \mu\text{g chl a l}^{-1}$ at Stn 9 near the Vistula river mouth) and low values were reported in the more saline open sea (less than $10 \mu\text{g C l}^{-1}$ and $1 \mu\text{g chl a l}^{-1}$). In average, concentrations were higher in the Gotland

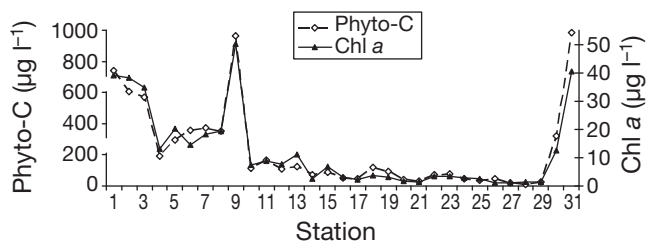


Fig. 3. Variations in phytoplankton C biomass (Phyto-C) and chl a concentrations among the stations

Sea than in the Bornholm Sea (Table 5), even when only offshore areas were considered. Over the whole data set, the average value of chl a was $12.19 \pm 13.98 \mu\text{g l}^{-1}$ and the chlide a concentration was only 3.1 (± 1.5)% of the total chl a (chl a + chlide a) concentration.

Relationships among group-specific chl a, marker pigments and Phyto-C

When the average of values from all stations was taken, the composition derived from the CHEMTAX analysis was in reasonable agreement with that derived from microscopic observations for dominant phytoplankton groups (dinoflagellates, cryptophytes and diatoms represented 50, 32 and 8% of the total C and 44, 33 and 19% of the chl a, respectively) (Fig. 4a,b). For the minor phytoplankton groups, the comparison between chl a and C biomass distributions was much more difficult due to the presence of unidentified small flagellates, which may represent several minor

Table 5. Mean ± SD phytoplankton biomass (wet wt), carbon biomass and chl a in 2 different regions (all units are $\mu\text{g l}^{-1}$)

Period	Gotland Sea			Bornholm Sea			Source
	Biomass	Carbon	chl a	Biomass	Carbon	chl a	
April 2005	3304 ± 2813	316 ± 294	17.49 ± 15.18	600 ± 894	67 ± 87	2.13 ± 1.02	Present study Wasmund et al. (2000)
March–May 1993–1997	1666		4.00	930		3.75	

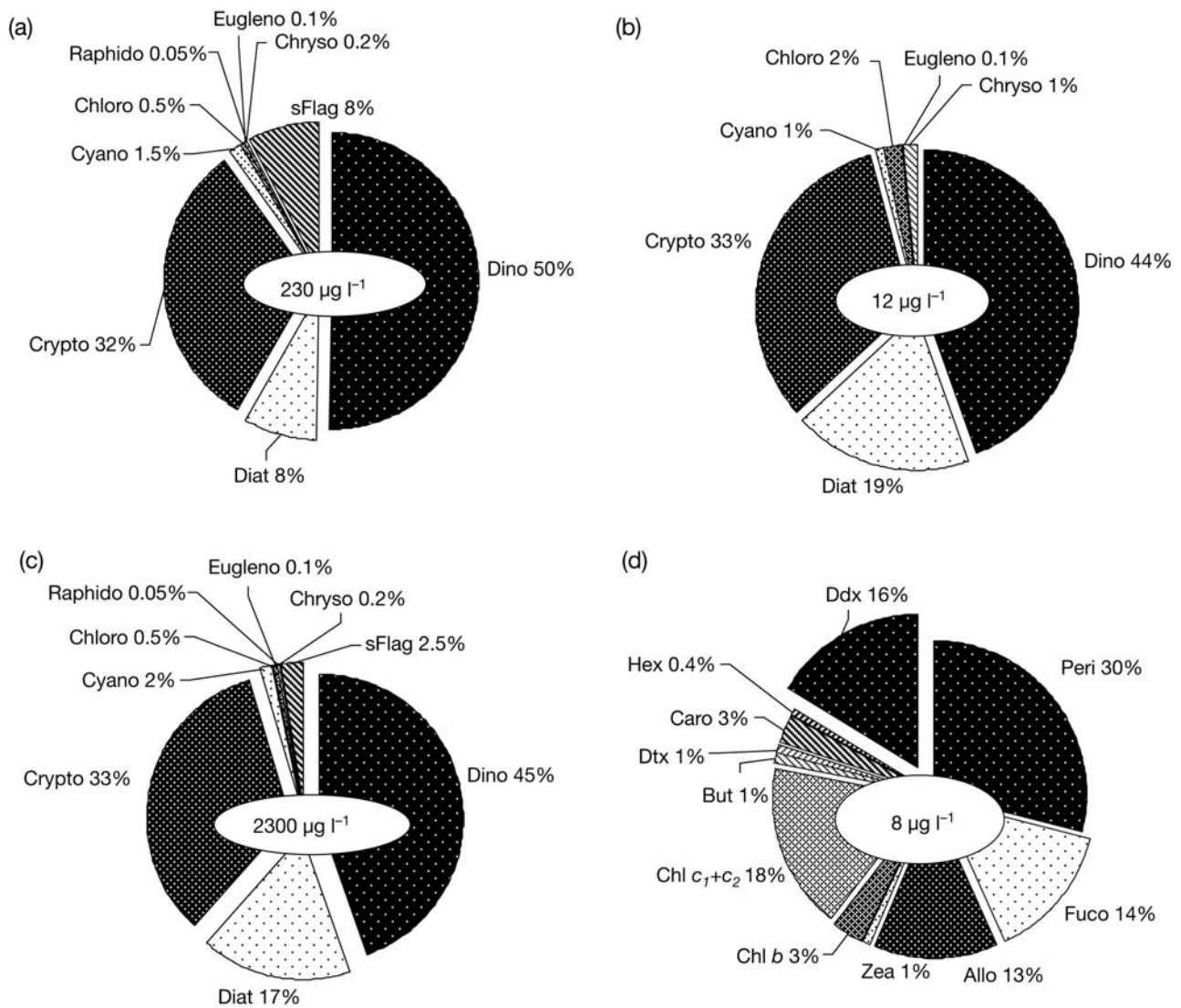


Fig. 4. Phytoplankton classification based on percentages of total (a) carbon biomass (b) chl *a* obtained from CHEMTAX analysis, (c) biomass (wet wt) and (d) accessory pigments. Numbers at the center of the pie graphs correspond to average absolute values of each variable at all stations. Abbreviations defined in Table 1

groups. The cumulative contribution of minor groups (Chloro, Chryso, Cyano, Eugleno) to the total C and chl *a* amounted to ~2.3% and 4%, respectively (Fig. 4a,b). Group composition of phytoplankton according to wet wt was mainly different for diatoms and small flagellates (Fig. 4c).

The average concentration of Peri, the unambiguous marker pigment of dinoflagellates, was $2.3 \pm 3.6 \mu\text{g l}^{-1}$, with values ranging from 0.03 to $14.5 \mu\text{g l}^{-1}$. Peri represented 30% of the sum of the measured accessory pigments. Other main contributors were chl c_1+c_2 ($1.4 \pm 1.9 \mu\text{g l}^{-1}$), Ddx ($1.3 \pm 1.9 \mu\text{g l}^{-1}$), Fuco ($1.2 \pm 1.3 \mu\text{g l}^{-1}$), Allo ($1.0 \pm 1.4 \mu\text{g l}^{-1}$) (Fig. 4d). The sum of the contributions by other minor pigments represented less than 9.5%.

Peri concentration was significantly correlated with Ddx and chl c_1+c_2 ($r^2 = 0.98$ and 0.92 , respectively, $p < 0.05$, Fig. 5a,b), the latter 2 showing almost equal concentrations (slope of the regression equal to 0.99, Fig. 5c) at all sampling stations except Stns 1 and 2, located in the Gdańsk Bight. Ddx and chl c_1+c_2 concentrations were also statistically correlated with Fuco ($r^2 = 0.56$, and $r^2 = 0.60$, respectively, $p < 0.05$, Fig. 5d,e), although the correlations were weaker than with Peri. Concentrations of the less dominant pigments, chl *b* and Zea, covaried among the stations ($r^2 = 0.89$, $p < 0.05$, Fig. 5f).

The variations of phytoplankton composition resulting from microscopy-derived carbon biomass, from

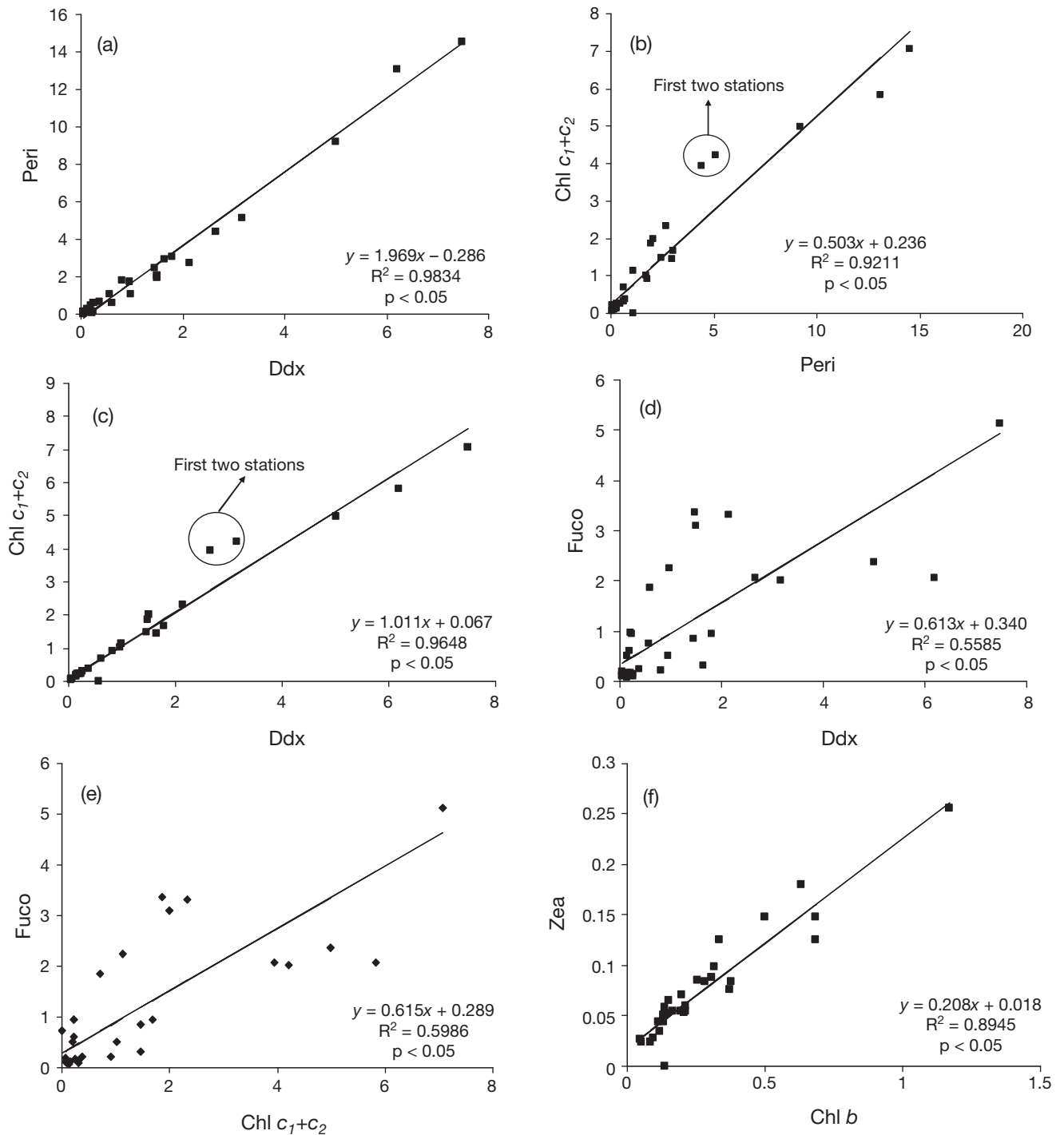


Fig. 5. Model II linear regressions (and corresponding parameters) computed for selected pairs of accessory pigments (units are $\mu\text{g l}^{-1}$). Abbreviations defined in Table 1

CHEMTAX and from the individual marker pigment composition were similar among the stations (Fig. 6). The highest C biomass, total accessory pigment and chl a concentrations were noted at the Bight of Gdańsk (Stns 1 through 11 and 31), where dino-

flagellates and cryptophytes were dominant. These groups were still dominant at the central Polish coasts (Stns 20, 21 and 29). However, in the Pomeranian Bight (Stns 22 through 25) the contribution of dinoflagellates, and thus of Peri, to the total carbon biomass and total

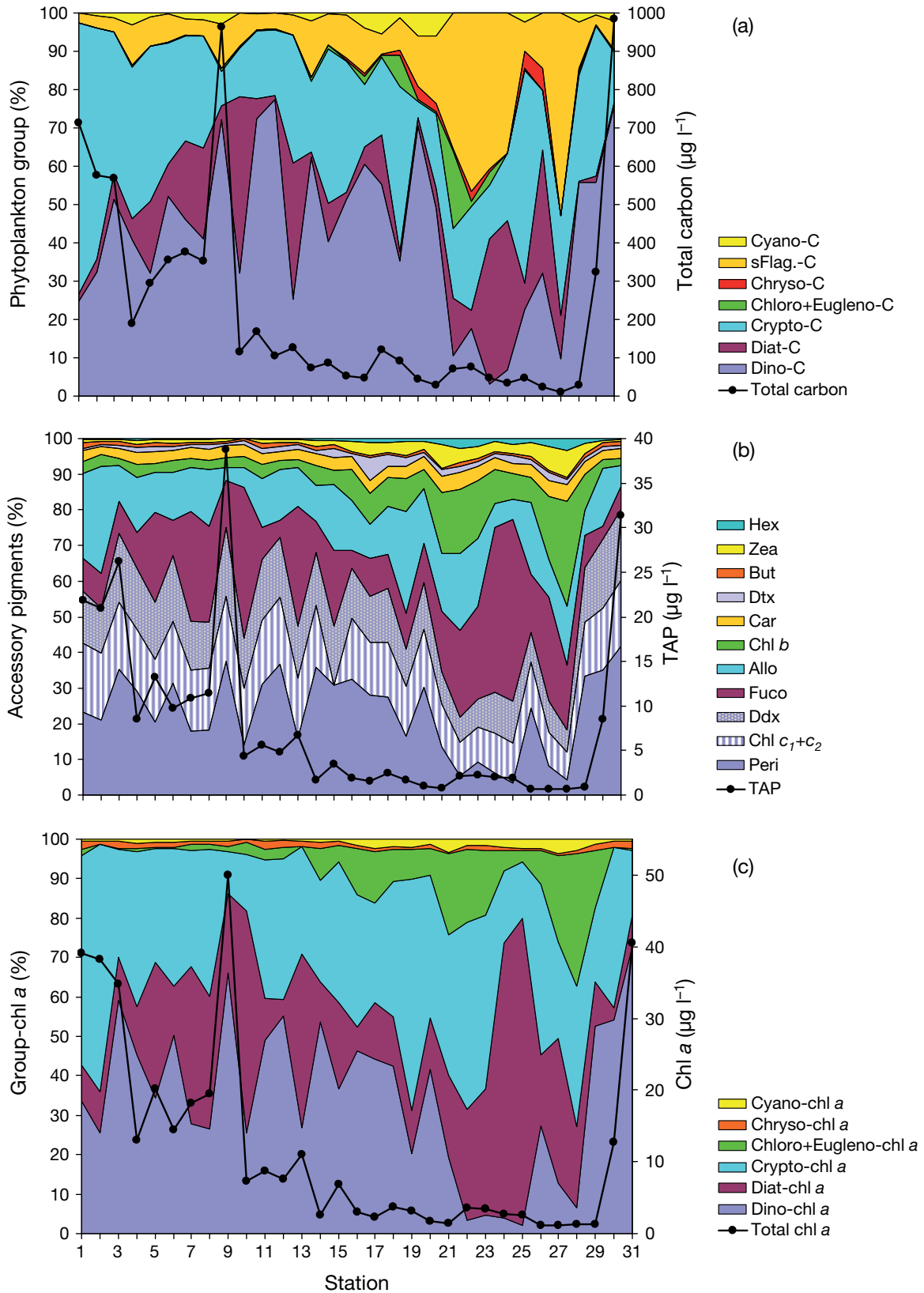


Fig. 6. Percentage contribution of (a) phytoplankton groups to the total carbon, (b) pigments to the total accessory pigments (TAP) and (c) CHEMTAX-allocated chl a of phytoplankton groups to the total chl a. Note that cryptophytes also contain the ciliate *Mesodinium rubrum*. Abbreviations defined in Table 1

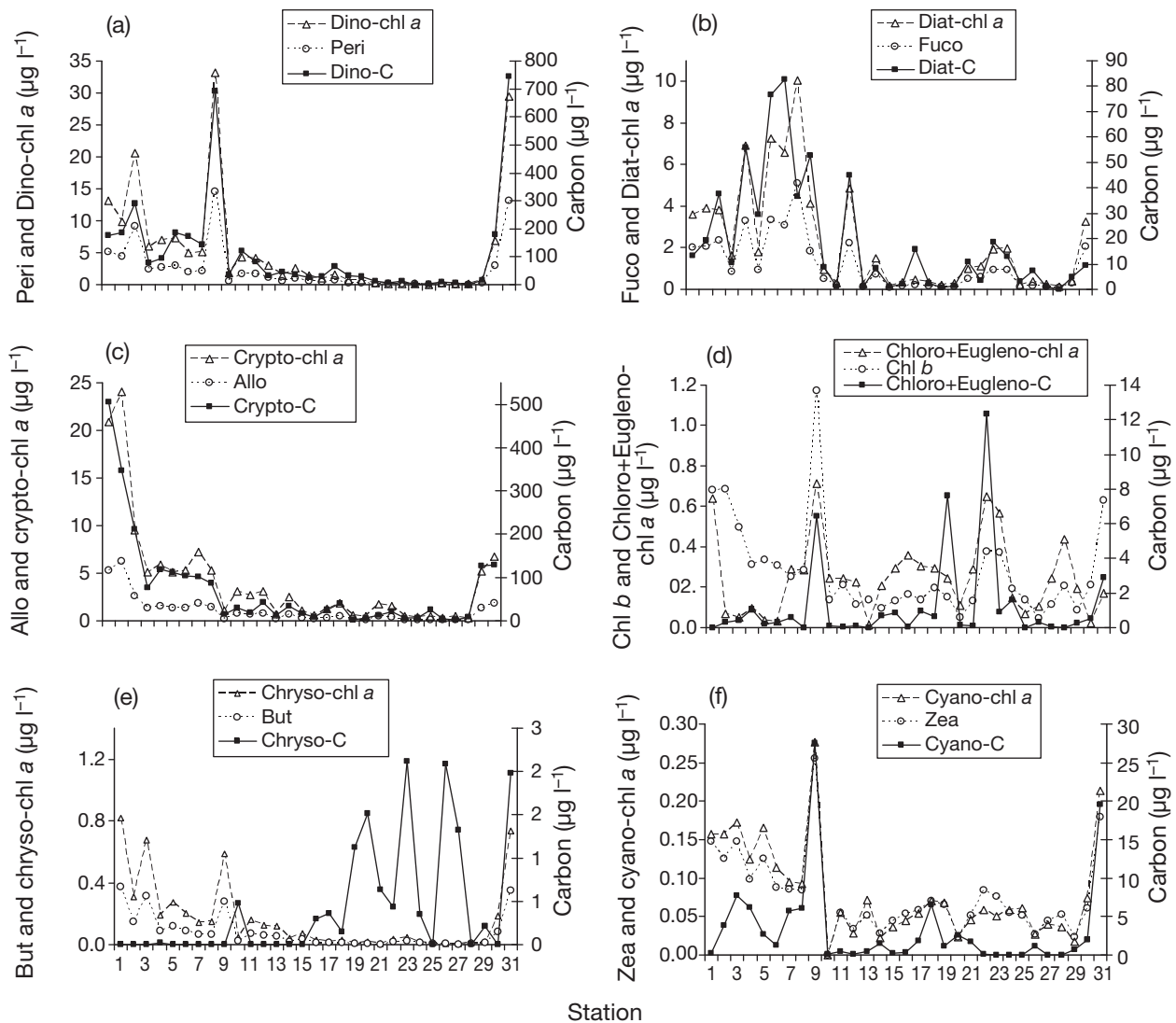


Fig. 7. Distribution of CHEMTAX-allocated chl *a* of phytoplankton groups together with the corresponding marker pigment and carbon concentrations (all units are $\mu\text{g l}^{-1}$). Abbreviations defined in Table 1

accessory pigment concentration, respectively, were considerably reduced, while diatom, small flagellates and chlorophyte contributions increased simultaneously with Fuco and chl *b* contributions (Fig. 6).

In terms of spatial distribution (Fig. 7a–c), C and chl *a* concentrations covaried with each other for major phytoplankton groups (dinoflagellates, diatoms and cryptophytes). A significant correlation was found between Dino-C and Peri (Dino-C = $50.49\text{Peri} - 2.22$, $r^2 = 0.94$, $p < 0.05$), as well as between Dino-C and CHEMTAX-allocated Dino-chl *a* ($r^2 = 0.93$, $p < 0.05$). Similarly, Diat-C was significantly correlated with Fuco (Diat-C = $22.33\text{Fuco} - 7.16$, $r^2 = 0.63$, $p < 0.05$) and Diat-chl *a* ($r^2 = 0.70$, $p < 0.05$). However, at some stations (e.g. Stns 1, 2, 9 and 31), CHEMTAX appeared to have overestimated Diat-chl *a* (Fig. 7b).

The correlation between Dino-C and Ddx ($r^2 = 0.90$, $p < 0.05$) was much higher than between Diat-C and Ddx ($r^2 = 0.10$, $p < 0.05$) (not shown). This was in accordance with the decrease in Ddx:chl *a* ratio for diatoms and its increase for dinoflagellates between the first input and the final output ratios of Matrix 1 (Table 2, not shown in the graph). These observations suggested that Ddx is a good marker pigment of dinoflagellates rather than of diatoms in the present investigation. Similarly, chl c_1+c_2 was much more correlated with Dino-C ($r^2 = 0.82$, $p < 0.05$) than with Diat-C ($r^2 = 0.13$, $p < 0.05$) and sFlag-C ($r^2 = 0.50$, $p < 0.05$).

C biomass of minor phytoplankton groups was either unrelated or poorly related with their marker pigments or with chl *a* allocated by CHEMTAX (Fig. 7d–f). CHEMTAX-allocated chl *a* of Chloro was poorly corre-

lated with C biomass of this group ($r^2 = 0.2$, $p < 0.05$) while there was no correlation between Eugleno-C and Eugleno-chl *a* ($p > 0.05$) (Fig. 7d). Since chrysophytes did not include But-containing pelagophytes, no correlation was observed between Chryso-C and Chryso-chl *a* (Fig. 7e). Cyano-C showed a rather good correlation with Zea ($r^2 = 0.66$, $p < 0.05$) and Cyano-chl *a* ($r^2 = 0.64$, $p < 0.05$), mainly due to a good fit at Stns 9 and 31 (Fig. 7f). While the correlation between marker pigments Zea and chl *b* was significant ($r^2 = 0.89$, $p < 0.05$), the correlation between Cyano-C (which contain Zea) and Chloro+Eugleno-C (which contain chl *b* and small amounts of Zea) was not significant ($r^2 = 0.06$, $p > 0.18$).

Crypto-C (including the phototrophic ciliate *Mesodinium rubrum*) and Allo were significantly correlated (Crypto-C = $80.29\text{Allo} - 10.98$, $r^2 = 0.91$, $p < 0.05$). Carbon biomass of *M. rubrum* was well above that of free-living cryptophytes present in the environment. These ciliates contained $89 \pm 17\%$ of Crypto-C ($95 \pm 9\%$ of the cryptophyte wet wt) at the first 2 stations (Fig. 8). High biomass of alloxanthin-containing organisms at these stations could also be deduced from pigment ratios. As previously noted, the chl c_1+c_2 :Ddx ratio was much higher at Stns 1 and 2 than at all other stations (Fig. 5c). Given that Peri and Ddx were closely associated with Dino-C, and also with chl c_1+c_2 , the excess of chl c_1+c_2 (chl c_2 is also a marker pigment of Crypto) at these stations could be attributed to cryptophytes found as endosymbionts in the environment.

C:chl *a* ratio and irradiance

Phyto-C was significantly correlated with chl *a* ($r^2 = 0.95$, $n = 31$, $p < 0.05$, Fig. 9a) and the average C:chl *a* ratio was equal to 20 ± 7 . The average C:chl *a* ratio was lower for diatoms (9 ± 7) than for dinoflagellates ($30 \pm$

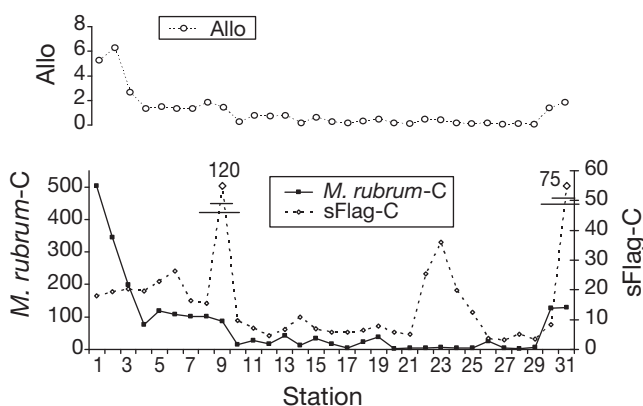


Fig. 8. Alloxanthin (Allo) concentrations and the distribution of the carbon biomass of small flagellates (sFlag) and *Mesodinium rubrum* (all units are $\mu\text{g l}^{-1}$)

17) and for cryptophytes (17 ± 9) (Fig. 9b–d). The use of different equations (see 'Materials and methods') resulted in a doubling of the ratio of both diatoms and cryptophytes. When these 2 parallel increases were taken into account, the C:chl *a* ratio of total phytoplankton became 28 ± 10 . The average C:chl *a* ratio considerably differs from the estimated slope of the regression for Cyanophytes as well as for the other groups (Chloro, Eugleno, Chryso and sFlag) (Fig. 9e,f). In fact, the significant linear relationship observed for each of these 2 groups is mainly driven by a couple of anomalous high values.

There was no significant correlation between C:chl *a* ratio and the average irradiance in the surface mixed layer either for all data (Fig. 10) or for clear sky samples ($y = 0.609 \ln(x) + 19.176$, $r^2 = 0.003$, $p > 0.83$). These ratios were lower than the optimal C:chl *a* ratios found using the model of Blackford et al. (2004). When C:chl *a* ratios of major phytoplankton groups, dinoflagellates, diatoms and cryptophytes, were separately regressed with I_{SML} , correlations were not significant either ($r^2 = 0.034$, $p = 0.319$; $r^2 = 0.001$, $p = 0.967$; $r^2 = 0.001$, $p = 0.886$, respectively).

DISCUSSION

Comparison of CHEMTAX results with different input ratio matrices

Latasa (2007) reported that the successive runs of CHEMTAX made the pigment ratios converge to the actual pigment:chl *a* ratios for most phytoplankton groups, greatly improving biomass estimates. In agreement with this statement, in the present study, Fuco:chl *a* for diatoms and Peri:chl *a* for dinoflagellates converged at the end of 10 successive runs, with a result of almost the same chl *a* allocations to these groups (Fig. 2). Similarly, the Allo:chl *a* ratio of cryptophytes, varying by a factor of 1.5 from Matrix 1 to Matrix 4 (Appendix 1A), converged after the first run. However, it remained unvaried, even after 10 successive runs, for the very high initial ratios of Matrix 5 (11- to 17-fold higher than in Matrices 1 through 4). Latasa (2007) also noted that if a minor group has only a single and unshared marker pigment, chl *a* concentrations could be correctly quantified, but if it shares marker pigments with major groups, CHEMTAX might not produce reliable results. A strict comparison of our results with Latasa's (2007) findings was not possible since all minor groups shared pigments with either another minor group (e.g. cyanophytes shared Zea with the other minor group chlorophytes) or with a major group (chrysophytes shared Ddx with the major groups, diatoms and dinoflagellates) in this work.

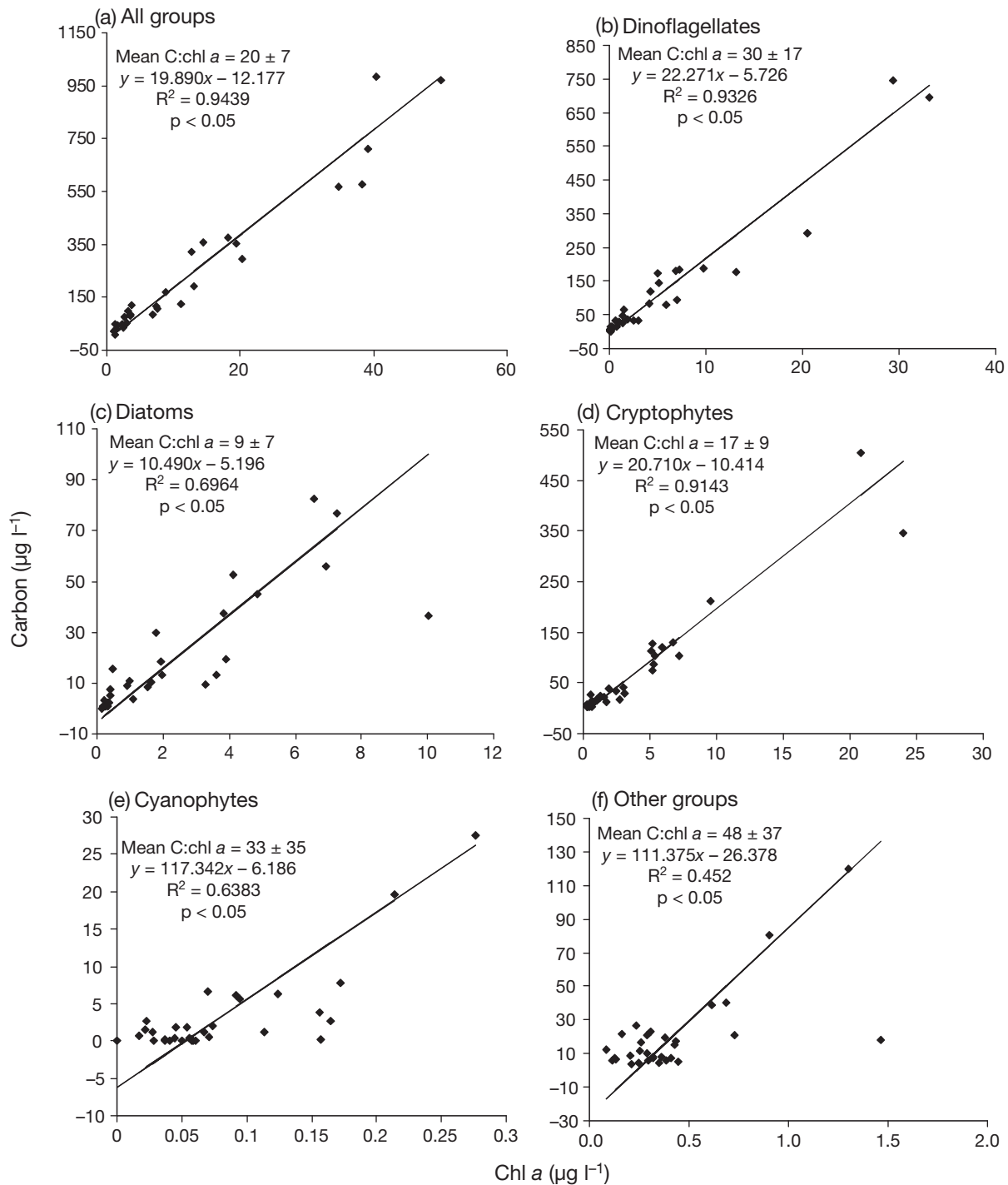


Fig. 9. Model II linear regressions of carbon versus chl *a* for (a) total phytoplankton, (b) dinoflagellates, (c) diatoms, (d) cryptophytes (including *Mesodinium rubrum*), (e) cyanophytes and (f) other groups (Chloro, Chryso, Eugleno, sFlag; see Table 1)

None of the pigment:chl *a* ratios of minor groups converged at the end of 10 successive runs in the present study, partially supporting the finding of Latasa (2007) about the minor groups.

Mackey et al. (1996, p. 268) reported that, 'care should also be taken when selecting what pigments to

use in the ratio matrix. Pigments that are present in nearly all phytoplankton are unlikely to give much useful information, while the use of pigments such as diadinoxanthin, which is converted rapidly to diatoxanthin in the light (Demers et al. 1991), or pigments which have wildly different abundances in different

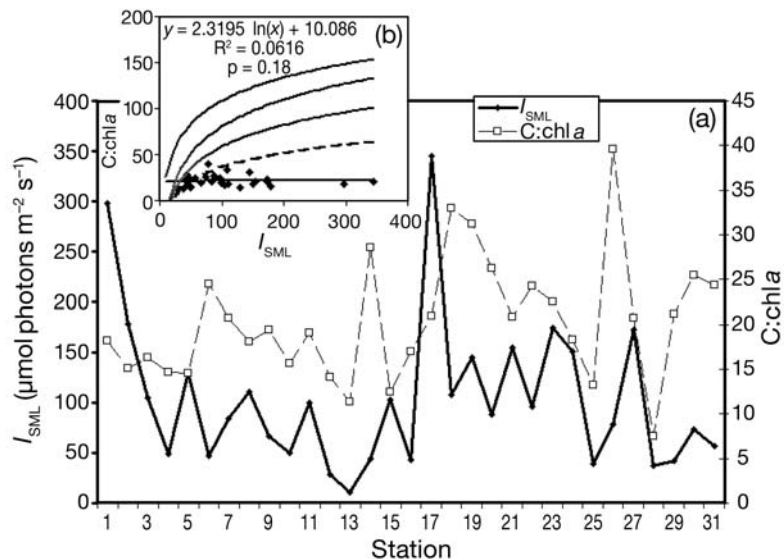


Fig. 10. (a) Variations in irradiance in the surface mixed layer (I_{SML}) and C:chl *a* ratios at the sampling stations. (b) Observed and model-derived C:chl *a* ratios under different irradiances. The black line and symbols (◆) represent the ratios observed in the present study. Grey lines show optimal C:chl *a* ratios derived from models of Blackford et al. (2004) and Geider et al. (1997). Each grey line, top to bottom, refers to a maximum assimilation rate of 1, 2 and 3 d^{-1} respectively. Ratios were constrained by maximum 150 and minimum 13. Dashed line was derived using the regression equation of Llewellyn et al. (2005) ($y = -0.0217 \ln(x) + 0.1353$, $r^2 = 0.91$, $p < 0.0001$ for 2001). The unit ' $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ ' was converted to ' W m^{-2} ' assuming $1 \text{ W m}^{-2} = 1.89 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ as in Llewellyn et al. (2005)

species within a class are also likely to give poor results.' When the widely shared chl c_1+c_2 , Caro and Dtx were included in the ratio matrices, concentrations of major groups were not affected but some minor groups were influenced. Euglenophytes were overestimated when these 3 pigments were used. Ddx was not excluded from any of the tested input ratio matrices even though it is a shared pigment by many phytoplankton groups, since in the present study, Ddx was very well correlated with Peri and Dino-C.

Phyto-C, Phyto-chl *a* and marker pigment relationship

The dominance of dinoflagellates during the study period at almost all stations was evidenced by all 3 methods used here to determine the phytoplankton composition: (1) the use of the sole marker pigments (Peri, Ddx, chl c_1+c_2), (2) the CHEMTAX analysis and (3) the microscopic examination (Fig. 4).

In many previous investigations performed in different seas, the chl *a* allocation to the dinoflagellates by CHEMTAX was not well correlated with the microscopic carbon evaluation of this class (Irigoiien et al. 2004, Llewellyn et al. 2005, Rodriguez et al. 2006).

This could be due to the presence of heterotrophic/mixotrophic dinoflagellates as well as of unusual pigment contents (in Fuco, Allo, Hex, chl *b*) of some dinoflagellates (Sournia et al. 1992, Johnsen & Sakshaug 1993, Jeffrey & Vesik 1997, Meyer-Harms & Pollehne 1998). By contrast, in the present investigation, significant positive correlations between Dino-C and Dino-chl *a*—as well as between Dino-C and the marker pigments Peri, Ddx and chl c_1+c_2 —were noteworthy (except chl c_1+c_2 at Stns 1 and 2). This must be due to the dominance of Peri-containing autotrophic dinoflagellates (Table 4). Good correlations between Dino-C and Peri, and also between Dino-C and Ddx, were reported previously in the Baltic Sea, in the Gulf of Bothnia and the Gulf of Finland (Piippola & Kononen 1995, Wänstrand & Snoeijs 2006).

According to CHEMTAX and microscopy results, the second important phytoplankton group contributing to the total chl *a* and C biomass was cryptophytes when the kleptoplastidic ciliate *Mesodinium rubrum* was included in that group (Figs. 4a–c, 6a,c). This ciliate was a significant source of Phyto-C and chl *a* in the Bight of Gdańsk, especially at Stns 1 and 2 (Fig. 8). *M. rubrum* was also previously reported to be an important phototrophic ciliate in the Gdańsk Basin (Witek 1998, Rychert 2004) and was noted to reach bloom concentrations (up to $\sim 200 \mu\text{g C l}^{-1}$) generally in April–July and October in the southern Baltic proper and Danish estuaries (Witek 1998, Henriksen et al. 2002, Schlüter & Møhlenberg 2003, Wasmund et al. 2005). Although not mentioned, *M. rubrum* could also be the explanation for the observed high Allo concentration while cryptophyte biomass was low in the northern Baltic Sea (Wänstrand & Snoeijs 2006). Crypto-C was significantly correlated with Allo ($r^2 = 0.91$) and with Crypto-chl *a* ($r^2 = 0.91$).

Diat-C values were also well correlated with Fuco and CHEMTAX-allocated chl *a* concentrations ($r^2 = 0.63$ and $r^2 = 0.70$, $p < 0.05$). However, at a few stations (e.g. Stns 9 and 31), diatoms seemed to be overestimated by CHEMTAX, probably due to the dominance of Fuco-containing small flagellates (e.g. chrysophytes) at these stations (Figs. 7b & 8). It was previously reported that large-sized diatoms had lower cellular chl *a* content with respect to small-sized individuals and thus were omitted in the comparison studies of biomass and pigment analyses

(Schlüter & Møhlenberg 2003 and references therein). In the present study, the discrepancy between Diat-C and Diat-chl *a* at the few stations mentioned above cannot be due to differences in cellular pigment content originating from distinct size composition of diatoms, because small-sized diatom species such as *Thalassiosira baltica* (Grunow) Ostefeld and/or *Skeletonema costatum* (Grev.) Cleve were dominant among the stations (Table 4).

The spatial distribution of phytoplankton groups containing chl *b* (chlorophytes and euglenophytes) or Zea (cyanophytes) generally differed according to the estimator of their abundance—the respective individual marker pigments chl *b* or Zea, CHEMTAX-allocated chl *a*, or C estimated from biovolume (Figs. 6 & 7d,f). Nonetheless, the contribution of these groups to the total accessory pigment and total carbon was low (Fig. 4a,d).

Spatial distribution of Phyto-C and marker pigments

It is helpful to know dominant phytoplankton groups, species and their bloom dynamics in a study region when estimating C biomass from pigment composition or from light absorption characteristics. In the present study, the dominant phytoplankton species were typical bloom-forming species of the region (Table 4). A spring dinoflagellate bloom, dominated by *Peridiniella catenata*, is common for the southern Baltic Sea according to long-term monitoring studies of the region (HELCOM 1996, Wasmund et al. 1998, Stoń et al. 2002, Gasiunaite et al. 2005). *P. catenata*-dominated blooms were also observed in the northern parts of the Baltic Sea, the Gulf of Finland and the Gulf of Riga in spring (Gasiunaite et al. 2005). According to these investigations (op. cit.), the biomass level of the bloom is generally higher in the southeastern Baltic Sea than in the southwestern part. Similarly, phytoplankton biomass and chl *a* concentration were several-fold higher in the eastern Baltic than in the western Baltic in the present investigation (Table 5). Such a difference in Phyto-C and chl *a* concentrations between the 2 areas could partially be due to the time lag between bloom occurrences as well. It was previously reported that the spring bloom initiates and ends earlier in the western part of the southern Baltic Sea than in the east (Wasmund et al. 1998, Gasiunaite et al. 2005). Moreover, it was noted that the bloom was mainly composed of diatoms in the southwestern Baltic and of dinoflagellates in the southeast (Wasmund et al. 1998, Gasiunaite et al. 2005). In the present study as well, diatoms and small flagellates were dominant at the 4 western-most stations (Stns 22 to 25), where the total C biomass was quite low.

The correlation between chlorophyte-euglenophyte and cyanophyte abundances suggested by the strong correlation between chl *b* and Zea (Fig. 5f) was not actually observed with microscopy ($p > 0.18$ and $r^2 = 0.06$ for the correlation between Chloro+Eugleno-C and Cyano-C). It could be due to possible inclusion of some chlorophytes into small flagellates at some stations, such as Stns 9 and 31 (Figs. 7d,f & 8) and/or contribution of picoplanktonic cyanobacteria to the C biomass of cyanophytes, which was not considered in this study.

C:chl *a* ratio

The average C:chl *a* ratio of total phytoplankton was 20 ± 7 in the present investigation. If the equations providing the highest estimation of C for diatoms and cryptophytes are taken into account, this ratio could be interpreted as 28 ± 10 (see also 'Results'). This ratio varies highly in nature, depending on light and nutrient status of phytoplankton species and groups (Cullen 1982). Harrison et al. (1977) reported variation from 20 to 60 in healthy cells. According to the modelling study of Cloern et al. (1995), which also compared the results of phytoplankton cultures under different light, temperature and nutrient conditions (a total of 219 published measurements), C:chl *a* ratios ranged between ~18 and ~333. The ratio found in the present study is at the lower end of the ranges reported previously (Table 6). It is not possible to estimate exact C values of phytoplankton species using C:V conversion equations, due to variations in this ratio depending on phytoplankton species, growth conditions and cell sizes (Verity et al. 1992, Montagnes et al. 1994, Llewellyn & Gibb 2000). For instance, C biomass of the diatom *Actinocyclus subtilis* was found to be 5 times higher with elemental C analysis than its C estimated from V using the equation of Strathmann (1967) (Llewellyn & Gibb 2000, Llewellyn et al. 2005). In addition, a 2 to 4-fold difference in C cell quotas was reported for the same laboratory cultures with remarkably similar V measured using the same C measurement technique, with an elemental C analyser (Verity et al. 1992, Montagnes et al. 1994). An inverse relationship between the cell V (~10 to $10^4 \mu\text{m}^3$) and the cellular C content was reported in previous studies (i.e. big cells have lower C density than small cells) (Verity et al. 1992, Llewellyn & Gibb 2000). C:V conversion equations, which were used in the present study and established by Strathmann (1967), Verity et al. (1992), Montagnes et al. (1994), Menden-Deuer & Lessard (2000) and Montagnes & Franklin (2001), involve the range of cell V found in our investigation (10 to $10^5 \mu\text{m}^3$). High cellular chl *a* content of dominant phytoplankton species could be an alternative hypothesis for low C:chl *a* ratios found in the re-

Table 6. C:chl *a* ratios of total phytoplankton, dinoflagellates (Dino) and diatoms (Diat) in different environments and in cultures, given as mean (\pm SD) and/or range. (–) Not given. Carbon estimation is from cell volumes using equations of (1) Strathmann (1967); (2) Verity et al. (1992); (3) Mullin et al. (1966); (4) Putt & Stoecker (1989); (5) Søndergaard et al. 1991; (6) Montagnes et al. (1994). In the laboratory cultures, carbon estimation was made by a CHN analyzer

Region	Total	Dino	Diat	C-estimation	Source
Southern Baltic	20 \pm 7	30 \pm 17	9 \pm 7	(1, 2)	Present study
Mesocosm, Isefjord, Denmark	–	6	50–100	(1, 3)	Havskum et al. (2004)
English Channel	23–78	666	10–48	(1, 2)	Llewellyn et al. (2005)
Iberian Poleward Current, Spain	161–303	32	79–250	(1, 2, 4)	Rodríguez et al. (2006)
Mesocosm, Denmark and Norway	24	–	–	(3, 5)	Schlüter & Havskum (1997)
Southern Ocean	45–90	–	–	(6)	Abraham et al. (2000)
Laboratory cultures		106	34	CHN	Chan (1980)
Laboratory cultures		365–4800	91–1749	CHN	Eker-Develi et al. (2006b)
Laboratory cultures		84 \pm 49,	18 \pm 4,	CHN	Geider et al. (1997) and
		21–143	14–28		references therein
Laboratory cultures			18–336	CHN	Laws & Bannister (1980)
Laboratory cultures			20–800	CHN	Sakshaug et al. (1989)

gion. Based on a 5 yr sampling study, Wasmund et al. (2001) reported that the peak chl *a* concentration (57 $\mu\text{g l}^{-1}$) was found during a dinoflagellate bloom in the Lithuanian coastal waters in May 1993. Phytoplankton biomass (wet wt) corresponding to this peak chl *a* concentration was 16 100 $\mu\text{g l}^{-1}$ and 93% of this biomass was formed by the dinoflagellate *Peridiniella catenata* (I. Olenina pers. comm.), which was also the dominant species in the present investigation in the Gotland Sea. If this biomass is roughly converted to C biomass by multiplying by 0.11 (Mullin et al. 1966), Phyto-C is found as 1770 $\mu\text{g l}^{-1}$, resulting in a C:chl *a* ratio of 31, which is similar to our C:chl *a* ratio for dinoflagellates (Table 6). Using the same C conversion factor (0.11) in April 1999 in the Bight of Gdańsk, Stoń et al. (2002) found that the C:chl *a* ratio was even lower (\sim 22) for the wet wt 7898 $\mu\text{g l}^{-1}$ and chl *a* \sim 40 $\mu\text{g l}^{-1}$. In that study as well, *P. catenata* was the dominant species, forming \sim 97% of the total wet wt.

It is known that nutrient limitation promotes an increase in the C:chl *a* ratio (Harrison et al. 1977, Laws & Bannister 1980, Sakshaug et al. 1989, Eker-Develi et al. 2006b). Simultaneous data on nutrients, pigments and phytoplankton cell counting were not available for the present study. However, based on nutrient measurements performed by the Institute of Meteorology and Water Management (IMGW 2005) 1 to 3 wk prior to our sampling (i.e. 4 to 8 April 2005), and phytoplankton C biomass estimated in the present investigation, we assume that nutrient limitation could be an issue along the central Polish coast (\sim 0.1 μM nitrate, 0.45 μM phosphate and 8 μM silicate) and in the Pomeranian Bight ($>$ 16.0 μM nitrate, 0.3 μM phosphate, 34 μM silicate), while this was less probable in the southeastern Gotland Sea (0.75 μM , 0.6 μM , 15 μM) and in the Bight of Gdańsk (70.0 μM nitrate, 1.2 μM phosphate, 70 μM silicate). Relatively high ratios of C:chl *a* were ob-

served along the central Polish coast and in the Pomeranian Bight (Stns 18 through 27, Fig. 2), with respect to the Bight of Gdańsk (Stns 1 through 11 and 31), which could be related to nutrient distribution.

There was no clear relationship between C:chl *a* ratio and the average irradiance within the surface mixed layer in the present study (Fig. 10), whereas Llewellyn et al. (2005) observed a positive correlation between C:chl *a* ratios and log of I_{SML} in the English Channel among 2 yr of weekly samples collected on clear sky days. Our observed C:chl *a* ratios were lower than our optimal C:chl *a* ratios, which were found using the phytoplankton acclimation models of Geider et al. (1997) and Blackford et al. (2004) (Fig. 10). These models were used by Llewellyn et al. (2005) and their observed C:chl *a* ratios for assimilation rates 1 to 3 d^{-1} were also below the model-derived C:chl *a* ratios, but higher than our observed C:chl *a* ratios (Fig. 10). Obviously, such a formulation of the relationship between irradiance and C:chl *a* did not take into account the light history of phytoplankton cells, but was only an instantaneous representation. It is also worth noting that according to Geider et al. (1997), estimates of C:chl *a* ratios based on their model might not accurately reflect the ratios of field populations because of substantial inter-specific variability in maximum growth rates, maximum chl *a* contents and light absorption characteristics of phytoplankton.

As expected, the C:chl *a* ratio here was higher for dinoflagellates (30 \pm 17) than for diatoms (9 \pm 7 and 18 \pm 14 when equations of Strathmann 1967 and Montagnes & Franklin 2001 were used, respectively). This finding is in qualitative agreement with differences reported by Chan (1980) for diatom and dinoflagellate cultures, and by Llewellyn et al. (2005) for *in situ* observations. In contrast, Rodríguez et al. (2006) and Havskum et al. (2004) noted much higher C:chl *a* for

diatoms than for dinoflagellates (Table 6). They noted that several factors could have caused these Diat-C:Diat-chl *a* ratios to be higher than Dino-C:Dino-chl *a* ratios, such as (1) the decrease in chl *a* content of diatoms in the late growth phase; (2) the dominance of large-celled diatoms with lower cellular chl *a* density; (3) the underestimation of Dino-C by microscopy; or (4) the decrease in chl *a* content of diatoms with increased light intensities.

CONCLUSION

The use of CHEMTAX together with the measured concentrations of marker pigments was quite successful for deriving biomass distribution of dominant phytoplankton groups in the southern Baltic Sea. Phytoplankton species composition and spatial distribution obtained from microscopy were similar to what was reported in previous years (Gasiunaite et al. 2005, Wasmund et al. 2005). It was observed that Peri, Ddx and chl c_1+c_2 concentrations were closely related to the C biomass of the dominant phytoplankton group, dinoflagellates, at the majority of stations. Excluding the phototrophic ciliate *Mesodinium rubrum* from cryptophyte biomass led to an important disagreement between microscopic biomass estimation and CHEMTAX-allocated chl *a* with this phytoplankton group.

In a few cases, the diatom contribution was overestimated by CHEMTAX analysis, probably due to the contribution of some small flagellates containing fucoxanthin, indicating the necessity of simultaneous microscopic analysis with pigment measurements.

Chl *a* and C biomass of minor phytoplankton groups were either not correlated significantly ($p > 0.05$ for euglenophytes) or poorly correlated ($r^2 = 0.2$, $p < 0.05$ for chlorophytes). The possible reasons could be linked to uncertainties with CHEMTAX when dealing with minor pigments in low concentrations (e.g. chl *b*, Zea), the difficulties in classifying small flagellates by microscopy, and the possible influence of phototrophic grazers on some of these minor groups. Even though cyanophytes were also among the minor phytoplankton groups, there was a relatively good correlation between their marker pigment zeaxanthin and their C biomass, mainly due to 2 high values.

Successive runs of CHEMTAX using the output of pigment ratios from each run as the input for the next provided a convergence of ratios for only major phytoplankton groups. Accordingly, resulting Peri:chl *a* and Fuco:chl *a* ratios of dinoflagellates and diatoms were 0.452 ± 0.02 and 0.489 ± 0.03 , respectively, for our Baltic Sea samples. However, such a convergence was not observed in any of the pigment ratios of minor phytoplankton groups.

The estimated C:chl *a* ratio of total phytoplankton (20 ± 7) in the Baltic Sea was at the lower end of previously reported ratios, which might be a characteristic of dominant species in the study region during spring. However, possible changes in the C:V ratio with varying size and species compositions is an issue which must be cautiously considered when presenting these ratios. C:chl *a* ratio was increased to 28 ± 10 upon usage of different C:V conversion equations.

Acknowledgements. This work has been supported by the JRC-European Commission. We acknowledge colleagues of the IOPAS, the Captain and the crew of the RV 'Oceania' for their support during the measuring campaign; E Lysiak-Pastuszak and IMGW for providing us their nutrient data; I. Olenina (Centre of Marine Research, Taikos, Klaipeda, Lithuania) for supplying us with phytoplankton carbon biomass data from 1993; and Drs. A. S. Heiskanen and A. C. Cardoso from JRC Ispra for allowing us to use their microscope. We are grateful to M. Dowell for early comments on the manuscript and to J. Challis for English corrections.

LITERATURE CITED

- Abraham ER, Law CS, Boyd PW, Lavender SJ, Maldonado MT, Bowie AR (2000) Importance of stirring in the development of an iron-fertilized phytoplankton bloom. *Nature* 407:727–730
- Alvain S, Moulin C, Dandonneau Y, Bréon FM (2005) Remote sensing of phytoplankton groups in case 1 waters from global SeaWiFS imagery. *Deep-Sea Res I* 52:1989–2004
- Blackford JC, Allen JI, Gilbert FJ (2004) Ecosystem dynamics at six contrasting sites: a generic modelling study. *J Mar Syst* 52:191–215
- Bralewska JM, Witek Z (1995) Heterotrophic dinoflagellates in the ecosystem of the Gulf of Gdańsk. *Mar Ecol Prog Ser* 117:241–248
- Chan AT (1980) Comparative physiological study of marine diatoms and dinoflagellates in relation to irradiance and cell size. II. Relationship between photosynthesis, growth and carbon-chlorophyll *a* ratio. *J Phycol* 16:428–432
- Claustre H, Hooker SB, Van Heukelem L, Berthon JF and others (2004) An intercomparison of HPLC phytoplankton pigment methods using in situ samples: application to remote sensing and database activities. *Mar Chem* 85: 41–61
- Cloern JE, Grenz C, Videgar-Lucas L (1995) An empirical model of the phytoplankton chlorophyll:carbon ratio—the conversion factor between productivity and growth rate. *Limnol Oceanogr* 40:1313–1321
- Cullen JJ (1982) The deep chlorophyll maximum: comparing vertical profiles of chlorophyll *a*. *Can J Fish Aquat Sci* 39: 791–803
- Demers S, Roy S, Gagnon R, Vignault C (1991) Rapid light-induced changes in cell fluorescence and in xanthophyll-cycle pigments of *Alexandrium excavatum* (Dinophyceae) and *Thalassiosira pseudonana* (Bacillariophyceae): photoprotection mechanism. *Mar Ecol Prog Ser* 76:185–193
- Eker-Develi E, Kideys AE, Tugrul S (2006a) Role of Saharan dust on phytoplankton dynamics in the northeastern Mediterranean. *Mar Ecol Prog Ser* 314:61–75
- Eker-Develi E, Kideys AE, Tugrul S (2006b) Effect of nutrients on culture dynamics of marine phytoplankton. *Aquat Sci* 68:28–39

- Gasiunaite ZR, Cardoso AC, Heiskanen AS, Henriksen P and others (2005) Seasonality of coastal phytoplankton in the Baltic Sea: influence of salinity and eutrophication. *Estuar Coast Shelf Sci* 65:239–252
- Geider RJ, MacIntyre HL, Kana TM (1997) Dynamic model of phytoplankton growth and acclimation: responses of the balanced growth rate and the chlorophyll *a*:carbon ratio to light, nutrient-limitation and temperature. *Mar Ecol Prog Ser* 148:187–200
- Gustafson DE Jr, Stoecker DK, Johnson MD, Van Heukelem WF, Sneider K (2000) Cryptophyte algae are robbed of their organelles by the marine ciliate *Mesodinium rubrum*. *Nature* 405:1049–1052
- Harrison PJ, Conway HL, Holmes RW, Davis CO (1977) Marine Diatoms grown in chemostats under silicate or ammonium limitation. III. Cellular chemical composition and morphology of *Chaetoceros debilis*, *Skeletonema costatum*, and *Thalassiosira gravida*. *Mar Biol* 43:19–31
- Havskum H, Schlüter L, Scharek R, Berdalet E, Jacquet S (2004) Routine quantification of phytoplankton groups—microscopy or pigment analyses? *Mar Ecol Prog Ser* 273: 31–42
- HELCOM (1996) Third periodic assessment of the state of the marine environment of the Baltic Sea, 1989–1993. Background Document, Baltic Sea Environ Proc 64B
- Henriksen P, Riemann B, Kaas H, Sorensen HM, Sorensen HL (2002) Effects of nutrient-limitation and irradiance on marine phytoplankton pigments. *J Plankton Res* 24:835–858
- Hooker SB, Claustre H, Ras J, Van Heukelem L and others (2000) The first SeaWiFS HPLC analysis round-robin experiment (SeaHARRE-1). In: Hooker SB, Firestone ER (eds) NASA Tech Memo. 2000–206892, vol. 14. NASA Goddard Space Flight Center, Greenbelt, MD, p 1–42
- IMGW (2005) Monitoring cruise report of R/V Baltica in the Polish sector of the southern Baltic Sea on 4–8 April 2005. Institute of Meteorology and Water Management, Maritime Branch, Gdynia
- Irigoien X, Meyer B, Harris R, Harbour D (2004) Using HPLC pigment analysis to investigate phytoplankton taxonomy: the importance of knowing your species. *Helgol Mar Res* 58:77–82
- Janson S (2004) Molecular evidence that plastids in the toxin-producing dinoflagellate genus *Dinophysis* originate from the free-living cryptophyte *Teleaulax amphioxeia*. *Environ Microbiol* 6:1102–1106
- Jeffrey SW, Vesik M (1997) Introduction to marine phytoplankton and their pigment signatures. In: Jeffrey SW, Mantoura RFC, Wright SW (eds) *Phytoplankton pigments in oceanography: guidelines to modern methods*, UNESCO monographs on oceanographic methodology no. 10. UNESCO, Paris, p 37–85
- Johnsen G, Sakshaug E (1993) Bio-optical characteristics and photoadaptive responses in toxic and bloom-forming dinoflagellates *Gyrodinium aureolum*, *Gymnodinium galatheanum*, and two strains of *Prorocentrum minimum*. *J Phycol* 29:627–642
- Joint Global Ocean Flux Study (1994) Protocols for the joint global ocean flux study core measurements. Intergovernmental Oceanographic Commission. SCOR. Manual and Guides, UNESCO 29:91–96
- Kim JS, Jeong HJ (2004) Feeding by the heterotrophic dinoflagellates *Gyrodinium dominans* and *G. spirale* on the red-tide dinoflagellate *Prorocentrum minimum*. *Mar Ecol Prog Ser* 280:85–94
- Kovala PE, Larrance JD (1966) Computation of phytoplankton cell numbers, cell volumes, cell surface and plasma volume volume per litre from microscopical counts. *Univ Wash Publ Oceanogr* 38:1–21
- Kowalczyk P, Olszewski J, Darecki M, Kaczmarek S (2005) Empirical relationships between coloured dissolved organic matter (CDOM) absorption and apparent optical properties in Baltic Sea waters. *Int J Remote Sens* 26:345–370
- Latasa M (2007) Improving estimations of phytoplankton class abundances using CHEMTAX. *Mar Ecol Prog Ser* 329:13–21
- Laws EA, Bannister TT (1980) Nutrient- and light-limited growth of *Thalassiosira fluviatilis* in continuous culture with implications for phytoplankton growth in the ocean. *Limnol Oceanogr* 25:457–473
- Llewellyn CA, Gibb SW (2000) Intra-class variability in the carbon, pigment and biomineral content of prymnesiophytes and diatoms. *Mar Ecol Prog Ser* 193:33–44
- Llewellyn CA, Fishwick JR, Blackford JC (2005) Phytoplankton community assemblage in the English Channel: a comparison using chl *a* derived from HPLC-CHEMTAX and carbon derived from microscopy cell counts. *J Plankton Res* 27:103–119
- Mackey MD, Mackey DJ, Higgins HW, Wright SW (1996) CHEMTAX—a program for estimating class abundances from chemical markers: application to HPLC measurements of phytoplankton. *Mar Ecol Prog Ser* 144:265–283
- Mantoura RFC, Llewellyn CA (1983) The rapid determination of algal chlorophyll and carotenoid pigments and their breakdown products in natural waters by reverse-phase high performance chromatography. *Anal Chim Acta* 151: 297–314
- Menden-Deuer S, Lessard EJ (2000) Carbon to volume relationships for dinoflagellates, diatoms, and other protist plankton. *Limnol Oceanogr* 45:569–579
- Meyer-Harms B, Pollehne F (1998) Alloxanthin in *Dinophysis norvegica* (Dinophysiales, Dinophyceae) from the Baltic Sea. *J Phycol* 34:280–285
- Montagnes DJS, Franklin DJ (2001) Effect of temperature on diatom volume, growth rate, and carbon and nitrogen content: reconsidering some paradigms. *Limnol Oceanogr* 46:2008–2018
- Montagnes DJS, Berges JA, Harrison PJ, Taylor FJR (1994) Estimating carbon, nitrogen, protein, and chlorophyll *a* from volume in marine phytoplankton. *Limnol Oceanogr* 39:1044–1060
- Mouritsen L, Richardson K (2003) Vertical microscale patchiness in nano- and microplankton distribution in a stratified estuary. *J Plankton Res* 25:783–797
- Mullin MM, Sloan PR, Eppley PW (1966) Relationship between carbon content, cell volume, and area in phytoplankton. *Limnol Oceanogr* 11:307–311
- Olenina I, Hajdu S, Edler L, Andersson A and others (2006) Biovolumes and size-classes of phytoplankton in the Baltic Sea. *HELCOM Baltic Sea Environ Proc* 106
- Piippola S, Kononen K (1995) Pigment composition of phytoplankton in the Gulf of Bothnia and Gulf of Finland. *Aqua Fenn* 25:39–48
- Putt M, Stoecker DK (1989) An experimentally determined carbon:volume ratio for marine ‘oligotrichous’ ciliates from estuarine and coastal waters. *Limnol Oceanogr* 34: 1097–1103
- Rodríguez F, Varela M, Zapata M (2002) Phytoplankton assemblages in the Gerlache and Bransfield Straits (Antarctic Peninsula) determined by light microscopy and CHEMTAX analysis of HPLC pigment data. *Deep-Sea Res II* 49:723–747
- Rodríguez F, Garrido JL, Crespo BG, Arbones B, Figueiras FG (2006) Size-fractionated phytoplankton pigment groups in the NW Iberian upwelling system: impact of the Iberian Poleward Current. *Mar Ecol Prog Ser* 323:59–73

- Rychert K (2004) The size structure of the *Mesodinium rubrum* population in the Gdańsk Basin. *Oceanologia* 46:439–444
- Sakshaug E, Andresen K, Kiefer DA (1989) A steady state description of growth and light absorption in the marine planktonic diatom *Skeletonema costatum*. *Limnol Oceanogr* 34:198–205
- Schlüter L, Havskum H (1997) Phytoplankton pigments in relation to carbon content in phytoplankton communities. *Mar Ecol Prog Ser* 155:55–65
- Schlüter L, Møhlenberg F (2003) Detecting presence of phytoplankton groups with non-specific pigment signatures. *J Appl Phycol* 15:465–476
- Schlüter L, Møhlenberg F, Havskum H, Larsen S (2000) The use of phytoplankton pigments for identifying and quantifying phytoplankton groups in coastal areas: testing the influence of light and nutrients on pigment/chlorophyll a ratios. *Mar Ecol Prog Ser* 192:49–63
- Sokal RR, Rohlf FJ (2000) *Biometry: the principles and practice of statistics in biological research*, 3rd edn. WH Freeman & Co, New York
- Søndergaard M, Jensen LM, Ærtebjerg G (1991) Picoalgae in Danish coastal waters during summer stratification. *Mar Ecol Prog Ser* 79:139–149
- Sournia A, Belin C, Billard C, Martial C and others (1992) The repetitive and expanding occurrence of a green, bloom forming dinoflagellate (Dinophyceae) on the coasts of France. *Cryptogam Algal* 13:1–13
- Stoń J, Kosakowska A, Łotocka M, Łysiak-Pastuszek E (2002) Pigment composition in relation to phytoplankton community structure and nutrient content in the Baltic Sea. *Oceanologia* 44:419–437
- Strathmann RR (1967) Estimating the organic carbon content of phytoplankton from cell volume or plasma volume. *Limnol Oceanogr* 12:411–418
- Tester PA, Geesey ME, Guo C, Paerl HW, Millie DF (1995) Evaluating phytoplankton dynamics in the Newport River estuary (North Carolina, USA) by HPLC-derived pigment profiles. *Mar Ecol Prog Ser* 124:237–245
- Uitz J, Claustre H, Morel A, Hooker S (2006) Vertical distribution of phytoplankton communities in open ocean: an assessment based on surface chlorophyll. *J Geophys Res* 111:C08005
- Veldhuis MLW, Kraay GW (2004) Phytoplankton in the subtropical Atlantic Ocean: towards a better assessment of biomass and composition. *Deep-Sea Res I* 51:507–530
- Verity PG, Robertson CY, Tronzo CR, Andrews MG, Nelson JR, Sieracki ME (1992) Relationships between cell volume and the carbon and nitrogen content of marine photosynthetic nanoplankton. *Limnol Oceanogr* 37:1434–1446
- Vidussi F, Claustre H, Manca BB, Luchetta A, Marty JC (2001) Phytoplankton pigment distribution in relation to the upper thermocline circulation in the Eastern Mediterranean Sea during winter. *J Geophys Res* 106:19939–19956
- Wänstrand I, Snoeijs P (2006) Seasonal phytoplankton pigment dynamics in the northern Baltic Sea: Assessment by ships-of-opportunity. *Estuar Coast Shelf Sci* 66:135–146
- Wasmund N, Nausch G, Matthäus W (1998) Phytoplankton spring blooms in the southern Baltic Sea—spatio-temporal development and long-term trends. *J Plankton Res* 20:1099–1117
- Wasmund N, Nausch G, Postel L, Witk Z and others (2000) Trophic status of coastal and open areas of the south-eastern Baltic Sea based on nutrient and phytoplankton data from 1993–1997. *Meereswiss Ber Warnemünde* 38:1–83
- Wasmund N, Andrushaitis A, Łysiak-Pastuszek E, Müller-Karulis B and others (2001) Trophic status of the south-eastern Baltic Sea: A comparison of coastal and open areas. *Estuar Coast Shelf Sci* 53:849–864
- Wasmund N, Pollehne F, Postel L, Siegel H, Zettler M (2005) Biologische Zustandseinschätzung der Ostsee im Jahre 2004. *Meereswiss. Ber., No 64, Warnemünde*, p 1–78
- Witek M (1998) Annual changes of abundance and biomass of planktonic ciliates in the Gdańsk Basin, southern Baltic. *Int Rev Hydrobiol* 83:163–182
- Wright SW, Jeffrey SW, Mantoura FC, Llewellyn CA, Bjørnland T, Repeta D, Welschmeyer N (1991) Improved HPLC method for the analysis of chlorophylls and carotenoids from marine phytoplankton. *Mar Ecol Prog Ser* 77:183–196

Editorial responsibility: Howard Browman, Storebø, Norway

*Submitted: July 17, 2007; Accepted: November 4, 2007
Proofs received from author(s): April 7, 2008*