

Coenzyme Q₁₀ treatment reduces lipid peroxidation, inducible and endothelial nitric oxide synthases, and germ cell-specific apoptosis in a rat model of testicular ischemia/reperfusion injury

In this experimental study, we assessed the preventive effects of coenzyme Q₁₀ (CoQ₁₀) in a rat model of ischemia/reperfusion injury. The results of this study show that CoQ₁₀ administration before the reperfusion period of testicular torsion provides a significant decrease in testicular lipid peroxidation products and expressions of inducible nitric oxide synthase, endothelial nitric oxide synthase, and germ cell-specific apoptosis. (*Fertil Steril*[®] 2010;93:280–2. ©2010 by American Society for Reproductive Medicine.)

Testicular torsion is a urologic emergency that causes testicular injury and subfertility (1). It appears that the main pathophysiology of testicular torsion is ischemia/reperfusion (I/R) injury of the testis caused by the twisted spermatic cord and its release (2, 3). Although reperfusion is essential for the survival of ischemic tissue, there is good evidence that reperfusion itself causes additional cell injury.

Coenzyme Q₁₀ (CoQ₁₀) is an essential component for electron transport in oxidative phosphorylation of mitochondria (4, 5). It is a potent antioxidant, a membrane stabilizer, and cofactor in the production of adenosine triphosphate by oxidative phosphorylation. Coenzyme Q₁₀ has been widely applied in food supplements and cosmetics in Japan, the U.S., and many other countries. In recent years, the frequency of studies involving CoQ₁₀ has increased in both basic and clinical research areas

(4–7). Several studies have demonstrated the protective effect of CoQ₁₀ in various forms of tissue injury (4). However, no study has investigated the effect of CoQ₁₀ in testicular I/R injury.

The present study aimed to assess the effects of CoQ₁₀ administration on lipid peroxidation, inducible nitric oxide synthase (iNOS), endothelial nitric oxide synthase (eNOS), and germ cell-specific apoptosis in a rat model of testicular I/R injury.

The study included 21 adult male Wistar rats weighing 240–260 g. The rats were maintained on a 12-hour light/dark cycle after ethical committee on animal research approval. Institutional Review Board approval was obtained. The rats were divided into three groups of seven rats each. One group underwent 1 hour of testicular torsion, the second received pretreatment with CoQ₁₀ before detorsion, and the third underwent a sham operation. Surgery was done with the subject under ketamine anesthesia (single intraperitoneal 50 mg/kg dose). All surgical procedures were performed through standard ilioinguinal incisions (8). At the end of the experiments, bilateral orchiectomies were performed and the rats were killed by pentobarbital overdose (200 mg/kg) and bilateral thoracotomy.

In the torsion-detorsion (T-D) group, the gubernaculum was divided, and the testis was freed from its longitudinal and distal pole attachment to the epididymis. Torsion was created by rotating the left testis 720° clockwise and maintained by fixing the testis to the scrotum with a 4-0 silk suture placed through the tunica albuginea. After 1 hour of torsion, the testis was counter-rotated back to the natural position and reinserted into the scrotum. The wound was closed using 3-0 catgut suture. After detorsion for 4 hours, bilateral orchiectomies were performed.

In the group receiving CoQ₁₀ before detorsion, the same surgical procedure was done as in the T-D group, but CoQ₁₀ (10 mg/kg; Sigma Chemical Co., St. Louis, MO) was injected intraperitoneally for 30 minutes before detorsion. The administration mode and dose of CoQ₁₀ corresponded to those used in earlier experimental studies (7). After detorsion for 4 hours, bilateral orchiectomies were performed.

In the control group (sham group), a sham procedure was performed. This consisted of same procedure as in the T-D group,

Bulent Erol, M.D.^a

Murat Bozlu, M.D.^e

Volkan Hanci, M.D.^b

Husnu Tokgoz, M.D.^a

Sibel Bektas, M.D.^c

Gorkem Mungan, M.D.^d

Departments of ^aUrology, Zonguldak Karaelmas University

Faculty of Medicine, Zonguldak

^bAnesthesiology, Zonguldak Karaelmas University Faculty of

Medicine, Zonguldak

^cPathology, Zonguldak Karaelmas University Faculty of

Medicine, Zonguldak

^dBiochemistry, Zonguldak Karaelmas University Faculty of

Medicine, Zonguldak

^eDepartment of Urology, University of Mersin School of

Medicine, Mersin, Turkey

Received May 11, 2009; revised July 9, 2009; accepted July 10, 2009; published online August 14, 2009.

B.E. has nothing to disclose. M.B. has nothing to disclose. V.H. has nothing to disclose. H.T. has nothing to disclose. S.B. has nothing to disclose. G.M. has nothing to disclose.

Reprint requests: Murat Bozlu, M.D., Department of Urology, University of Mersin School of Medicine, Zeytinlibahce Caddesi, 33079-Mersin, Turkey (FAX: 90-324-337-4305; E-mail: muratbozlu@yahoo.com).

TABLE 1The values of MDA levels and iNOS, eNOS and APAF-1 expressions in ipsilateral and contralateral testes (mean \pm SD).

Group	MDA (μ mol/g protein)		iNOS (positive staining/tubule)		eNOS (positive staining/tubule)		APAF-1 (positive staining/tubule)	
	Ipsilateral	Contralateral	Ipsilateral	Contralateral	Ipsilateral	Contralateral	Ipsilateral	Contralateral
Torsion-detorsion ^a	1.99 \pm 0.45	0.38 \pm 0.12	5.88 \pm 2.01	1.15 \pm 0.85	6.55 \pm 3.08	1.12 \pm 0.54	25.62 \pm 6.39	10.15 \pm 1.22
CoQ ₁₀ ^b	0.71 \pm 0.21	0.4 \pm 0.18	2.57 \pm 0.78	1.13 \pm 0.75	3.28 \pm 0.95	1.16 \pm 0.62	11.00 \pm 1.15	9.95 \pm 1.18
Sham	0.42 \pm 0.15	0.39 \pm 0.1	1.12 \pm 0.92	1.15 \pm 0.95	1.18 \pm 0.42	1.14 \pm 0.7	9.28 \pm 0.95	9.75 \pm 1.15

Note: There were no statistically significant differences in the contralateral testes among the groups ($P > .05$).APAF-1 = apoptosis protease-activating factor 1; CoQ₁₀ = coenzyme Q₁₀; eNOS = endothelial nitric oxide synthase; iNOS = inducible nitric oxide synthase; MDA = malondialdehyde.^aVersus sham in the ipsilateral testes; $P < .001$.^bVersus torsion-detorsion in the ipsilateral testes; $P < .01$.

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except that after rotating the testis 720° clockwise, it was immediately relieved, and a 4-0 silk suture was placed through the tunica albuginea. Bilateral orchiectomies were performed after the procedure.

Testicular lipid peroxidation was estimated by the measurement of malondialdehyde (MDA) (9). High-performance liquid chromatographic (HPLC) analysis was performed using a Shimadzu HPLC system (Kyoto, Japan) with MDA kit (Immundiagnostik, Bensheim, Germany). Immunohistochemical evaluation of iNOS, eNOS, and germ cell-specific apoptosis were performed with iNOS Ab-1, eNOS Ab-1, and apoptosis protease-activating factor 1 (APAF-1) antibody (Lab Vision Corp., Neomarkers, CA), respectively (10). The number of stained germ cells were counted in 100 seminiferous tubules on circular cross-sections for each group.

All data are expressed as mean \pm SD. Analysis of variance was used for statistical analysis of the data among the groups. Multiple comparisons were made using Tukey procedure, with $P < .05$ considered to be statistically significant.

The values of testicular MDA levels and iNOS, eNOS, and APAF-1 expressions of each group are shown in Table 1. Compared with the sham group, testicular MDA levels and expressions of iNOS, eNOS, and APAF-1 obtained from the T-D group were significantly higher in the ipsilateral testes ($P < .001$). Ipsilateral testicular MDA levels and iNOS, eNOS, and APAF-1 expressions obtained from the CoQ₁₀ treatment group were significantly lower than those obtained from the T-D group ($P < .01$). All of the parameters of contralateral testes did not reveal any statistically significant differences among the groups ($P > .05$).

In agreement with earlier studies (8, 11, 12), we found that testicular T-D caused a significant increase in testicular lipid peroxidation products and expressions of iNOS and eNOS and germ cell-specific apoptosis. The results from the present study indicated that CoQ₁₀ administration before the reperfusion period of testicular torsion provided a significant decrease in testicular lipid peroxidation products and expressions of iNOS, eNOS, and germ cell-specific apoptosis.

Coenzyme Q₁₀ was first introduced as an ethical drug for heart failure patients in Japan and other nations. Coenzyme Q₁₀, which functions endogenously in the mitochondrial electron transport chain, can be ingested to scavenge free radicals and contribute to antioxidant defenses in vivo (5). In recent years, the role of CoQ₁₀ in disease prevention and treatment has been intensely investigated (4). The successful results of CoQ₁₀ administration in different organ systems led us to attempt such treatment with a model of testicular I/R injury. To our knowledge, the effect of CoQ₁₀ on testicular I/R injury has not been reported.

We demonstrated that administration of CoQ₁₀ treatment before reperfusion caused significant reduction in the level of testicular MDA. It has been demonstrated that CoQ₁₀ functions as an antioxidant, scavenging free radicals and inhibiting lipid peroxidation (5). The present data indicate that treatment with CoQ₁₀ prevents further increase in lipid peroxidation caused by I/R; therefore, it may protect the testis against oxidative damage.

The results of earlier studies suggest that NO has an important role in damage of the testis via I/R (8, 12). It has been investigated in several organs that after ischemia, superoxide is produced during the reperfusion phase which rapidly reacts with NO and forms

reactive nitrogen species peroxynitrite (2). Peroxynitrite initiates toxic oxidative reactions, directly inhibits mitochondrial respiratory enzymes, decreases cellular oxygen consumption, and inhibits membrane sodium transport. Coenzyme Q₁₀ can reverse endothelial dysfunction by preventing oxidative and nitrative stress and inflammation (5). It may act to scavenge oxidant species, thereby reducing oxidative stress and resulting in recoupling of NOS (13). We demonstrated that treatment with CoQ₁₀ significantly decreases the expressions of iNOS and eNOS. In the light of these data, one may anticipate that administering CoQ₁₀ before reperfusion would protect the testis against NO-related injury in I/R.

Turner et al. (11) reported that testicular T-D in the rat resulted in germ cell-specific apoptosis. In the present study, we showed that T-D caused a significant increase in APAF-1 expression in the testis. APAF-1 plays a central role in mitochondrial apoptosis (14). The release of cytochrome c from the mitochondria leads to recruitment of procaspase 9 and APAF-1 into a protein complex that subsequently activates caspase 9. Lysiak et al. (15) found that IR of the testis stimulated a mitochondrial- and caspase 9-dependent pathway to germ cell-specific apoptosis. To our knowledge, the role of CoQ₁₀ on germ cell-specific apoptosis of testis induced by I/R is undefined. Under our experimental conditions, we found that administering CoQ₁₀ before reperfusion caused a significant decrease in the testicular APAF-1 expression compared with that in the T-D group.

Recent investigations have emphasized the antiapoptotic effect of CoQ₁₀ and the protective effect of CoQ₁₀ against apoptotic changes in several tissues and cells (16, 17). Emerging evidence indicates that a central event of apoptosis is opening of the mito-

chondrial permeability transition pore. Opening of the permeability transition pore is responsible for disruption of the mitochondrial transmembrane electrochemical gradient and is accompanied by extrusion to cytoplasm of several molecules, including cytochrome c and APAF-1, responsible for caspase cascade activation. The antiapoptotic activity of CoQ₁₀ is mediated by hindering mitochondrial depolarization, cytochrome c release to cytoplasm, and activation of caspase 9 (16, 17). Coenzyme Q₁₀ may be endowed with antiapoptotic activity as a modulator of permeability transition pore opening. On the other hand, we demonstrated that CoQ₁₀ may confer protective effect against lipid peroxidation in testicular I/R injury, and germ cell-specific apoptosis may be suppressed by this antioxidant action of CoQ₁₀. However, this study is the first report in the literature to show that the apoptotic changes seen in germ cells were reduced by CoQ₁₀ in the rat model of testicular torsion.

To date, several enzymes and drugs have been studied to alleviate testicular damage in animal models of I/R injury (1, 10, 18). However, none have been tested in clinical trials, apart from cooling the scrotum. Coenzyme Q₁₀ was first introduced as a drug for heart failure patients; its present status in most countries is that of a compound aimed at improving cellular bioenergetics, counteracting oxidative stress, and slowing down some age-related pathologies; it is also used as a preventive support measure.

In conclusion, the use of CoQ₁₀ in humans in both earlier studies and clinically without significant side effects can make its potential use in testicular torsion more attractive. The results of the present experimental study show that administration of CoQ₁₀ may be a novel approach for the therapy of I/R injury of the testis.

REFERENCES

1. Visser AJ, Heyns CF. Testicular function after torsion of the spermatic cord. *BJU Int* 2003;92:200-3.
2. Cuzzocrea S, Riley DP, Caputi AP, Salvemini D. Antioxidant therapy: a new pharmacological approach in shock, inflammation, and ischemia/reperfusion injury. *Pharmacol Rev* 2001;53:135-59.
3. Lysiak JJ, Nguyen QA, Kirby JL, Turner TT. Ischemia-reperfusion of the murine testis stimulates the expression of proinflammatory cytokines and activation of c-jun N-terminal kinase in a pathway to E-selectin expression. *Biol Reprod* 2003;69:202-10.
4. Littarru GP, Tian L. Clinical aspects of coenzyme Q₁₀: an update. *Curr Opin Clin Nutr Metab Care* 2005;8:641-6.
5. Belardinelli R, Tian L, Littarru GP. Oxidative stress, endothelial function and coenzyme Q₁₀. *Biofactors* 2008;32:129-33.
6. Hatanaka J, Kimura Y, Lai-Fu Z, Onoue S, Yamada S. Physicochemical and pharmacokinetic characterization of water-soluble coenzyme Q₁₀ formulations. *Int J Pharm* 2008;363:112-7.
7. Miles MV. The uptake and distribution of coenzyme Q(10). *Mitochondrion* 2007;7S:72-7.
8. Bozlu M, Eskandari G, Cayan S, Canpolat B, Akbay E, Atik U. The effect of poly(adenosine diphosphate-ribose) polymerase inhibitors on biochemical changes in testicular ischemia-reperfusion injury. *J Urol* 2003;169:1870-3.
9. Esterbauer H, Schaur RJ, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med* 1991;11:81-128.
10. Yagmurdur H, Ayyildiz A, Karaguzel E, Akgul T, Ustun H, Germiyanoglu C. Propofol reduces nitric oxide-induced apoptosis in testicular ischemia-reperfusion injury by downregulating the expression of inducible nitric oxide synthase. *Acta Anaesthesiol Scand* 2008;52:350-7.
11. Turner TT, Tung KS, Tomomasa H, Wilson LW. Acute testicular ischemia results in germ cell-specific apoptosis in the rat. *Biol Reprod* 1997;57:1267-74.
12. Shirashi K, Naito K, Yoshida K. Nitric oxide promotes germ cell necrosis in the delayed phase after experimental testicular torsion of rat. *Biol Reprod* 2001;65:514-21.
13. Chew GT, Watts GF. Coenzyme Q₁₀ and diabetic endotheliopathy: oxidative stress and the "recoupling hypothesis." *QJM* 2004;97:537-48.
14. Zuo Y, Xiang B, Yang J, Sun X, Wang Y, Chang H, et al. Oxidative modification of caspase-9 facilitates its activation via disulfide-mediated interaction with APAF-1. *Cell Res* 2009;19:449-57.
15. Lysiak JJ, Zheng S, Woodson R, Turner TT. Caspase-9-dependent pathway to murine germ cell apoptosis: mediation by oxidative stress, BAX, and caspase 2. *Cell Tissue Res* 2007;328:411-9.
16. Witort E, Pattarino J, Papucci L, Schiavone N, Donnini M, Lapucci A, et al. Autologous lipofilling: coenzyme Q₁₀ can rescue adipocytes from stress-induced apoptotic death. *Plast Reconstr Surg* 2007;119:1191-9.
17. Wu KLH, Hsu C, Chan JYH. Impairment of the mitochondrial respiratory enzyme activity triggers sequential activation of apoptosis-inducing factor-dependent and caspase-dependent signaling pathways to induce apoptosis after spinal cord injury. *J Neurochem* 2007;101:1552-66.
18. Dokmeci D, Kanter M, Inan M, Aydogdu N, Basaran UN, Yalcin O, Turan FN. Protective effects of ibuprofen on testicular torsion/detorsion-induced ischemia/reperfusion injury in rats. *Arch Toxicol* 2007;81:655-63.